9/5430 80 Rec'd PCT/PTO 09 DEC1997 130,00-154

Attorney's Docket No. .

U 011415-0

CHAPTER II

IN THE UNITED STATES ELECTED OFFICE (EO/US)

PCT/AU96/00149

15 MARCH 1996

16 MARCH 1995

INTERNATIONAL APPLICATION NO INTERNATIONAL FILING DATE ANTIGEN COMPOSITION AGAINST MYCOPLASMA

PRIORITY CLAIMED

TITLE OF INVENTION

JOHN WALKER

APPLICANT(S)

2. ROGAN LEE

3. STEPHEN WILLIAM DOUGHTY

Box PCT

Commissioner of Patents and Trademarks

Washington, D.C. 20231

ATTENTION: EO/US

COMPLETION OF FILING REQUIREMENTS FOR INTERNATIONAL APPLICATION ENTERING U.S. NATIONAL STAGE IN U.S. ELECTED OFFICE (EO/US) UNDER 35 USC 371

(check and complete the applicable item, if applicable)

This replies to the Notice of Missing Requirements under 35 U.S.C. 371 and 37 CFR 1,495 (FORM PCT/DO/EO/905).

□ A copy of FORM PCT/DO/EO/905 accompanies this response.

WARNING: Where the items being submitted to complete the entry of the international application into the - national phase are subsequent to 30 months from the priority date the application is still considered to be in the international state and if mailing procedures are utilized to obtain a date the express mail procedure of 37 CFR 1.10 must be used (because international application papers are not covered by an ordinary certificate of mailing 37 CFR 108(2)(xi)

NOTE Documents and fees must be clearly identified as a submission to enter the national stage under 35 USC 371. Otherwise, the submission will be considered as being made under 35 USC 111 37 CFR 1.495(g).

CERTIFICATE UNDER 37 CFR 1.10

I hereby certify that this Completion of Filing Requirements and the papers indicated as being transmitted therewith as "EXPRESS MAIL POST OFFICE TO ADDRESSEE" Mailing Label Number __E152803780911S. addressed to the: Commissioner of Patents and Trademarks, Washington, D.C. 20231.

/82/1988 TH BLFE | 01000008 01713450 FRYSHS

NOTE Each paper referred to as enclosed herein has the number of the "Express Mail" mailing label placed thereon pnor to mailing, 37 CFR 1.16(b)

WARNING: Certificate of mailing (first class) or facsimile transmission procedures of 37 CFR 1.8 cannot be used to obtain a date of mailing or transmission for this correspondence

(Completion of Filing Requirements for International Application Entering U.S. Elected Office (EO/US) [13-19]—page 1 of 5)

DECLARATION OR OATH

- I.

 No onginal declaration or oath was filed. Enclosed is the original declaration or oath for this application.
 - NOTE: For surcharge fee for filing declaration after filing date complete item IV(3)
- NOTE: Acceptable minimums in the declaration in an ordinarily filed U.S. application for identification of the specification to which it applies are the name of the inventor and (1) senal number, (2) attorney docket number that was on the application as filed and the filing date, (3) title of the invention and filing date, (4) title of invention and reference to a specification that is attached to the declaration at the time of execution and filed with the declaration, or (5) title of invention and a statement by a registered attorney that the application filed in the PTO is the application which the inventor executed by signing the declaration. If the identification (4) is used it must be accompanied by a statement that the "attached" specification is a copy of the specification and any amendments thereto that were filed in the PTO to obtain the filing date. Such a statement must be a venfied statement if made by a person not registered to practice before the PTO. Notice of September 12, 1983 (1035 O.G. 3).
- NOTE: Another minimum found acceptable in the declaration is the filing date (i.e., date of express mail) and the express mail number, useful where the senal number is not yet known. But note the practice where the express mail deposit is a Saturday, Sunday or holiday within the District of Columbia. 37 CFR 1.10(c)
- NOTE: 37 CFR 1 41(a) points out that "Full names must be stated, including the family name and at least one given name without abbreviation together with any other given name or initial."

(complete (c) or (d), if applicable)

Atta	che	d ıs	а
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- (c) Statement by a registered attorney that the application filed in the PTO is the application that the inventor executed by signing the declaration.
- (d) Statement that the "attached" specification is a copy of the specification and any amendments thereto that were filed in the PTO to obtain the filing date.

AMENDMENT

(complete as applicable)

An amendment in accordance with 37 CFR § 1.121 is attached.

The attached amendment cancels claims ______ inclusive.

TRANSMITTAL OF ENGLISH TRANSLATION OF NON-ENGLISH LANGUAGE PAPERS

- III.
 Submitted herewith is a English translation of the non-English language international application papers as originally filed. It is requested that this translation be used as the copy for examination purposes in the PTO. (See 37 CFR 1.495(c))
 - NOTE: For fee for processing a non-English application, complete item IV(4)
 - NOTE. A non-English oath or declaration in the form provided or approved by the PTO need not be translated 37 CFR 1.69(b)
- NOTE: Unlike the filing of an ordinary non-English application (37 CFR 1.52(d)), the translation of an international application entering the U.S. national phase need not be verified. 37 CFR 1.495(c). If necessary, however, a verified translation may be required. 37 CFR 1.495(c). Moreover, if the English translation is filed within 30 months from the priority date, no processing fee is required.

FEES

IV.			
NOTE.	The fees for claims and surcharge fees listed below in items 1 and 2 are reduced a small entity status is established on or before the date the fee is paid. If a verified statement is filled within 2 months of the date of timely payment of fee paid will be refunded on request. 37 CFR 1.28(a).	the full	fee was paid, but
1. Fe	es for claims		
	each independent claim in excess of 3 (37 CFR 1.492(b))—\$78.00 small entity—\$39.00 each claim in excess of 20	\$	
_	(37 CFR 1.492(c))\$22.00; small entity\$11.00	\$	
	multiple dependent claims(s) (37 CFR 1.492(d))—\$250.00 small entity—\$125.00	\$	
2. Su	rcharge fees		
	surcharge set forth in 37 CFR 1.492(e) for accepting the declaration later than 30 months after the priority date in filing an application in the U.S. as a designated office—\$130.00; small entity—\$65.00	\$	
NOTE	The processing fee in the next item 3 below is not subject to a reduction for	or sma ll	entity status
3. ₺	processing fee set forth in 37 CFR 1.492(f) for acceptance of an English translation later than 30 months after the priority date—\$130.00	\$	130.00
	Total fees	\$	
	SMALL ENTITY STATUS		
v . 🗆	A verified statement that this filing is by a small entity		
NOTE	If an original verified statement and a refund request are filed within two mont of a fee, then the excess fee paid will be refunded on request. 37 CFR 1.2		e date of payment
	(check and complete applicable items)		
	is attached.		
	☐ A separate refund request accompanies this paper.		

EXTENSION OF TIME

(complete (a) or (b), as applicable)

VI.			
The proced CFR § 1.136	edings herein are foi (a) apply.	a patent application	on. Accordingly, the provisions of 37
(a) 🗌 A _l 37	oplicant petitions for CFR § 1.17(a)-(d),	an extension of tir for the total numbe	me, the fees for which are set out in r of months checked out below:
Exten	sion Fe	e for other than	Fee for
(mon	ths)	small entity	small entity
☐ one r		\$ 110.00	\$ 55.00
= ::	nonths	\$ 380.00	\$ 190.00
_	months months	\$ 900.00 \$1,400.00	\$ 450. 00 \$ 700.0 0
		¥ 2 ,100.00	V 700.00
If an addition	nal extension of time	Fee:	\$consider this a petition therefor.
ii dii deeliioi	di extension or time	is required, please	consider this a petition therefor.
	(check and c	omplete the next ite	em, if applicable)
□ Ar th	n extension for erefor of \$	months has a is deducted from	ready been secured, and the fee paid the total fee due for the total months
of	extension now requ	ested.	
Ex	ktension fee due witt	this request \$	
		or	
tic	onal petition is being	made to provide	erm is required. However, this condi- for the possibility that applicant has etition and fee for extension of time.
		TOTAL FEE DU	E
VII.			
The total for	ee due is:		
Con	pletion fee(s)	\$	
	nsion fee (if any)	\$	
	(,,	TOTAL FEE	DUE \$ 130.00
		PAYMENT OF FE	EES
VIII.			
⊠ Er	nclosed is a check in	n the amount of \$_	130.00
			mount of \$
	duplicate of this rec		
	should be itemized in suc		ar for which purpose the fees are paid. 37 CFR
·		ents for International Ap	plication Entering U.S. Elected Office (EO/US) [13-19]—page 4 of 5)

AUTHORIZATION TO CHARGE ADDITIONAL FEES

IX.		
WARN	ING: Accurately count claims, especial featra claims are authorized	ally multiple dependant claims, to avoid unexpected high charges
WARNI	ING: "Submission of the appropriate or petition for extension is filed.	extension fee under CFR 1.136(a) is to no avail unless a request " Notice of November 5, 1985 (1060 O.G. 27)
×	The Commissioner is hereb that may be required by t application to Account No.	y authorized to charge the following additional fees his paper and during the entire pendency of this 12-0425
		.92(a)(4) (filing fees)
	☐ 37 CFR 1.492(b), (c), a	nd (d) (presentation of extra claims)
NOTE.	set for response by the PTO in any n	nultiple dependent claims not paid on filing or on later presentation incelled by amendment prior to the expiration of the time period actice of fee deficiency (37 CFR 1.16(d)), it might be best not to discuss accept possibly when dealing with amendments after
	37 CFR 1.17 (application	on processing fees)
WARNI	should be made only with the kno	(d) deal with extensions of time under § 1 136(a) this authorization owledge that "Submission of the appropriate extension fee under tiless a request or petition for extension is filed " (Emphasis added) 60 O.G. 27)
	37 CFR 1.18 (issue fee a to 37 CFR 1.311(b).	at or before mailing of Notice of Allowance, pursuant
NOTE	Where an authorization to charge the of a Notice of Allowance, the issue fee of mailing the notice of allowance. 37	issue fee to a deposit account has been filed before the mailing will be automatically charged to the deposit account at the time 7 CFR 1.311(b)
NOTE:	of 37 CFR 1.28(b) (a) notification of c	of any change in loss of entitlement to small entity status must paying, or at the time of paying — . issue fee "From the wording thange of status must be made even if the fee is paid as "other ation is required if the change is to another small entity
	☐ 37 CFR 1.492(e) and/or	(f) surcharge fees for filing the declaration and/or
	an English translation of from the earliest-claime	of an international application later than 30 months
WARNII	VG: It would be wise to always check	
Reg. No.:	:	SIGNATURE OF ATTORNEY Clifford J. Mass
Tel. No.:	()	type or/print name of sito help tion No. 30,086 c/ollabas & PARK 25 nost Clat Streat
		P.O. Address Xev Fork, 27 10025 (212) 708-1890

77 Rec'd PCT/PTO 12SEP19S7 08/913430

Attorney's	Docket	No.	U	011	415	-0

CHAPTER !!

TRANSMITTAL LETTER TO THE UNITED STATES ELECTED OFFICE (EO/US)

(ENTRY INTO U.S. NATIONAL PHASE UNDER CHAPTER ID

INTERNATIONAL		
PCT/AU96/00149		149 15 MARCH 1996 16 MARCH 1999
TITLE OF INVEN	ITION	40 1211011 133
		ANTIGEN COMPOSITION AGAINST MYCOPLASMA
APPLICANT(S)	1.	JOHN WALKER
	2.	ROGAN LEE
Box PCT	3.	STEPHEN WILLIAM DOUGHTY
Assistant 6		

Assistant Commissioner for Patents

Washington D.C. 20231

ATTENTION: EO/US

NOTE: The completion of those filing requirements that can be made at a time later than 30 months from the priority date results from the Commissioner exercising his judgment under the authority granted under 35 USC 371(d). The filing receipt will show the actual date of receipt of the last item completing the entry into the national phase. See 37 CFR 1.491 which states: "An international application enters the national state when the applicant has filed the documents and fees required by 35 USC 371(c) within the periods set forth in § 1.494 and § 1.495."

WARNING: Where the items are those which can be submitted to complete the entry of the international application into the national phase are subsequent to 30 months from the priority date the application is still considered to be in the international state and if mailing procedures are utilized to obtain a date the express mail procedure of 37 CFR 1.10 must be used (since international application papers are not covered by an ordinary certificate of mailing - 37 CFR 1.8 (2) (xi)).

NOTE: Documents and fees must be clearly identified as a submission to enter the national state under 35 USC 371 otherwise the submission will be considered as being made under 35 USC 111. 37 CFR 1.494(f).

CERTIFICATION UNDER 37 CFR 1.10

I hereby certify that this Transmittal Letter and the papers indicated as being transmitted therewith is being deposited with the United States Postal Service on this date $\underbrace{SEPT}_{-} 12_{F} 1997_{-}$, in an envelope as "Express Mail Post Office to Addressee" Mailing Label Number $\underbrace{EH684275215}_{EH684275215}$, addressed to the: Assistant Commissioner for Patents, Washington, D.C. 20231.

GERALDINE MARTI

(type or print name of person mailing paper)

Signature of person mailing paper

NOTE: Each paper or fee referred to as enclosed herein has the number of the "Express Mail" mailing label placed thereon prior to mailing. 37 CFR 1.16(b).

WARNING: Certificate of mailing (first class) or facsimile transmission procedures of 37 CFR 1.8 cannot be used to obtain a date of mailing or transmission for this correspondence.

(Transmittal Letter to the United States Elected Office (EO/US) [13-18]-page 1 of 8)

- I. Applicant herewith submits to the United States Elected Office (EO/US) the following items under 35 U.S.C. 371:
 - a. X This express request to immediately begin national examination procedures (35 U.S.C. 371(f)).
 - b. X The U.S. National Fee (35 U.S.C. 371(c)(1)) and other fees (37 CFR 1.492) as indicated below:

2. Fees

CLAIMS FEE	(1) FOR	(2) NUMBER FILED	(3) NUMBER EXTRA	(4) RATE	(5) CALCULA- TIONS	
□ *	TOTAL CLAIMS					-
		43 -20=	23	× \$22.00 =	\$ 506.00	NOT PAID AT
	INDEPENDENT CLAIMS					THIS TIME
		8 -3=	5	×\$80.00=	400.00	NOT PAID AT
	MULTIPLE DEPE	ENDENT CLAIM(S) (if	applicable)	+\$260.00		THIS TIME
BASIC FEE*	AUTHORITY	AS INTERNATIONAL I				
	in § 1.482 ha	ternational preliminar as been paid on the i	y examination fed international appl	e as set forth ication to the		
	sta	d the international prates that the criteria viousness) and indus	of novelty, invent	ive step (non-		
	Ar	ticle 33(1) to (4) have nims presented in the	been satisfied for	r all the		
	na	tional stage (37 CFR d the above requiren	1.492(a)(4))	\$96.00		
	1.4	192(a)(1))S NOT INTERNATION		\$700 on		
	EXAMINATIO	N AUTHORITY emational preliminar				
	in § 1.482 ha	s been paid to the U search fee as set for	.S. PTO, and pay	ment of an		
	☐ has	s been paid (37 CFR s not been paid (37 C	1.492(a)(2))	\$770.00		
	□ wh	ere a search report of been prepared by the	on the internation	al application		
	the	Japanese Patent Of 92(a)(5))	fice (37 CFR	\$910.00		
	· · · · · · · · · · · · · · · · · · ·		Total of abov	e Calculations	=	
SMALL	Reduction by 1/2	for filing by small en		<u>- </u>	1,040.00	
ENTITY	must be filed also	o. (note 37 CFR 1.9, 1	.27, 1.28)	į.	-	
				Subtotal		•
	F 1				1,040.00	
	Fee for recording CFR 1.21(h)). (See COVER SHEET".	the enclosed assigni Item 13 below). See	ment document \$ attached "ASSIG	40.00 (37 NMENT		
OTAL			Total F	ees enclosed \$	1,040.00	

*See att	ched Preliminary Amendment Reducing the Number of Claims.	
	X A check in the amount of 1,040.0 to cover the above fees is enclos	ed.
	Please charge Account No in the amount of \$ A duplicate copy of this sheet is enclosed.	
**WARNIN	"To avoid abandonment of the application the applicant shall furnish to the United States Pa and Trademark Office not later than the expiration of 30 months from the priority date: " the basic national fee (see § 1.492(a)). The 30-month time limit may not be extended." 37 (§ 1.495(b).	1 (2)
WARNING	If the translation of the international application and/or the oath or declaration have not be submitted by the applicant within thirty (30) months from the priority date, such requirements a be met within a time period set by the Office. 37 CFR § 1.495(b)(2). The payment of the surchaset forth in § 1.492(e) is required as a condition for accepting the oath or declaration later thirty (30) months after the priority date. The payment of the processing fee set forth in § 1.492 is required for acceptance of an English translation later than thirty (30) months after the priority date. Failure to comply with these requirements will result in abandonment of the application. provisions of § 1.136 apply to the period which is set. Notice of January 3, 1993, 1147 O.G. to 40.	may arge han 92(f) ority The
3. 🙀	copy of the International application as filed (35 U.S.C. 371(c)(2)):	
ap "7 ac co de ap no	ion 1.495 (b) was amended to require that the basic national fee and a copy of the internation of the internation of the priority date to avoid abandonment of the priority date of the priority date of the priority date. In accordance with PCT Rule 47.1, that notice shall be accepted by practed offices as conclusive evidence that the communication has duly taken place. Thus, if the international stage, the applicant normally need only check to be sure the from the International Bureau has been received and then pay the basic national fee by 30 months the priority date." Notice of January 7, 1993, 1147 O.G. 29 to 40, at 35-36. See item 14c belong the priority date."	ent. e in the all the the the
	is transmitted herewith.	
	. is not required, as the application was filed with the United Star Receiving Office.	tes
	. 🖾 has been transmitted	
	 i.	
	ii. by applicant on (date)	
4. □ X	translation of the International application into the English language 5 U.S.C. 371(c)(2)):	
	is transmitted herewith.	
	. 👿 is not required as the application was filed in English.	
	was previously transmitted by applicant on (date)	
	will follow.	

5.	X	An (35	nend 5 U.S	Iments to the claims of the International application under PCT Article 19 S.C. 371(c)(3)):
NO	f C S	nnd co priority do so submit an a m	ontinu date will r that endn	of January 7, 1993 points out that 37 CFR § 1.495(a) was amended to clarify the existing uning practice that PCT Article 19 amendments must be submitted by 30 months from the e and this deadline may not be extended. The Notice further advises that: "The failure to not result in loss of the subject matter of the PCT Article 19 amendments. Applicant may subject matter in a preliminary amendment filed under section 1.121. In many cases, filing ment under section 1.121 is preferable since grammatical or idiomatic errors may be 1147 O.G. 29-40, at 36.
		a.		are transmitted herewith.
		b.		have been transmitted
			i.	☐ by the International Bureau. Date of mailing of the amendment (from form PCT/1B/308):
			ii.	☐ by applicant on (date)
		c.	Ķ	have not been transmitted as
			i.	applicant chose not to make amendments under PCT Article 19. Date of mailing of Search Report (from form PCT/ISA/210.):
			ii.	☐ the time limit for the submission of amendments has not yet expired. The amendments or a statement that amendments have not been made will be transmitted before the expiration of the time limit under PCT Rule 46.1.
6.				lation of the amendments to the claims under PCT Article 19 s.C. 371(c)(3)):
		a.		is transmitted herewith.
		b.		is not required as the amendments were made in the English language.
		c.		has not been transmitted for reasons indicated at point 5c above.
7.	□ x	A c	ору	of the international examination report (PCT/IPEA/409)
			X	is transmitted herewith.
				is not required as the application was filed with the United States Receiv-Office.
8.		Anr	ex(e	es) to the international preliminary examination report
		a.		is/are transmitted herewith.
		b.	□ Red	is/are not required as the application was filed with the United States ceiving Office.
9.		A tr	ansl	ation of the annexes to the international preliminary examination report
		a.		is transmitted herewith.
		b.		is not required as the annexes are in the English language.

10. 🛭	An oath or declaration of the inventor (35 U.S.C. 371(c)(4)) complying with 35 U.S.C. 115
	a. was previously submitted by applicant on (date)
	b. is submitted herewith, and such oath or declaration
	i. is attached to the application.
	ii.
	iii. X will follow.
II. Other of	document(s) or information included:
11. 🙀	An International Search Report (PCT/ISA/210) or Declaration under PCT Article 17(2)(a):
	a. 🛚 is transmitted herewith.
	b. has been transmitted by the International Bureau. Date of mailing (from form PCT/IB/308):
	c. \square is not required, as the application was searched by the United States International Searching Authority.
	d.
	e. has been submitted by applicant on (date)
12. 🙀	An Information Disclosure Statement under 37 CFR 1.97 and 1.98:
	a. is transmitted herewith.
	Also transmitted herewith is/are:
	☐ Form PTO-1449.
	☐ Copies of citations listed.
	b.
	c. was previously submitted by applicant on (date)
13. 🗆	An assignment document is transmitted herewith for recording.
	A separate "COVER SHEET FOR ASSIGNMENT (DOCUMENT) ACCOMPANYING NEW PATENT APPLICATION" or FORM PTO 1595 is also attached.

14. 🛚	A E	dditional documents:
	a.	☐ Copy of request (PCT/RO/101)
	b.	International Publication No. <u>WO 96/</u> 28472
		i. X Specification, claims and drawing
		ii. Front page only
	c.	☐ Preliminary amendment (37 CFR § 1.121)
	d.	
		FORM PCT/IPEA/402
	_	
15. ⋤	₫ Tr	e above checked items are being transmitted
	a.	before 30 months from any claimed priority date.
	b.	after 30 months.
16.		ertain requirements under 35 U.S.C. 371 were previously submitted by the plicant on, namely:
	α _L	, namely.
		AUTHORIZATION TO CHARGE ADDITIONAL FEES
WARNII	NG: A	Accurately count claims, especially multiple dependant claims, to avoid unexpected high charges featra claims are authorized.
	Ç X	The Commissioner is hereby authorized to charge the following additional fees that may be required by this paper and during the entire pendency of this application to Account No. $\underline{12-0425}$
		□x 37 CFR 1.492(a)(1), (2), (3), and (4) (filing fees)
WARNII	VG: E	Because failure to pay the national fee within 30 months without extension (37 CFR § 1.495(b)(2)) esults in abandonment of the application, it would be best to always check the above box.
		☐ 37 CFR 1.492(b), (c) and (d) (presentation of extra claims)
NOTE:	must set for	ise additional fees for excess or multiple dependent claims not paid on filing or on later presentation only be paid or these claims cancelled by amendment prior to the expiration of the time period response by the PTO in any notice of fee deficiency (37 CFR 1.492(d)), it might be best not to rize the PTO to charge additional claim fees, except possible when dealing with amendments after ction.

(Transmittal Letter to the United States Elected Office (EO/US) [13-18]—page 7 of 8)

0,086
et 23

PTO/PST Rec'd 2 OCT 1997-

08/9/3430

PATENT THE UNITED STATES PATENT AND TRADEMARK OFFICE

In re application of: J

John WALKER, et al

Serial No.:

08/913,430

Group No: ---

Filed:

OCT 0 2 1997

September 12, 1997

Examiner:

For:

ANTIGEN COMPOSITION AGAINST MYCOPLASMA

Attorney Docket No.:

U-011415-0

Commissioner Patents and Trademarks Washington, DC 20231

AMENDMENT

Sir:

Prior to an examination of this application on the merits, please amend the application as follows:

IN THE CLAIMS:

Please cancel claims 1 - 43 and replace with the following new claims.

CERTIFICATE OF MAILING (37 CFR 1.8a)

I hereby certify that this paper (along with any paper referred to as being attached or enclosed) is being deposited with the United States Postal on the date shown below with sufficient postage as first class mail in an envelope addressed to the: Commissioner of Patents and Trademarks, Washington, DC 20231

CLIFFORD MASS

Type or print hame of person mailing paper)

Date: September 30, 1997

(Signature of person mailing paper)

, and

20

44. A putative protective antigen against a <u>Mycoplasma</u>, prepared by a method including

providing

a sample of a Mycoplasma;

an antibody probe including at least antibody against a Mycoplasma produced by a method including

providing a biological sample taken a short time after an immune animal has been challenged with a <u>Mycoplasma</u> or <u>Mycoplasma</u> extract taken from the infection site or an area of a lesion or an area close to the infection site or lesion;

isolating cells from the biological sample; culturing cells in vitro in suitable culture medium; and harvesting antibodies produced from said cells;

probing the <u>Mycoplasma</u> sample with the antibody probe to detect at least one antigen; and

isolating the antigen detected.

- 45. A putative protective antigen against <u>Mycoplasma hyopneumoniae</u>, or related infections, selected from the group of antigens having approximate molecular weights of 110-114, 90-94, 72-75, 60-64, 52-54 and 46-48 kilodaltons (kD), mutants, derivatives and fragments thereof.
- 46. A putative protective antigen according to claim 45 wherein the antigen in the 72-75 kD region contains the following N-terminal amino acid sequence:

 AGXLQKNSLLEEVWYLAL
 - 47. A putative protective antigen according to claim 46 further including one or more of the following internal amino acid sequences:

30 AKNFDFAPSIQGYKKIAHEL
NLKPEQILQLLG
LLKAEXNKXIEEINTXLDN

48. A putative protective antigen according to claim 45 wherein the antigen in the 60-64 kD region contains the following N-terminal amino acid sequence:

MKLAKLLKGFX(N/L)(M/V)IK, or ADP(F/I)(R/E)Y(V/A)PQG(Q/A)X(M/N)VG

5

49. A putative protective antigen according to claim 45 wherein the antigen in the 52-54 kD region contains the following N-terminal amino acid sequence:

AGXWAKETTKEEKS

10 50. A putative protective antigen according to claim 49 further including one or more of the following internal amino acid sequences:

AWVTADGTVN AIVTADGTVNDNKPNQWVRKY

15 51. A putative protective antigen according to claim 45 wherein the antigen in the 46-48 kD region contains the following N-terminal amino acid sequence:

AGXGQTESGSTSDSKPQAETLKHKV

52. A putative protective antigen according to claim 51 further including one or more of the following internal amino acid sequences:

TIYKPDKVLGKVAVEVLRVLIAKKNKASR AEQAITKLKLEGFDTQ KNSQNKIIDLSPEG

25 53. An isolated nucleic acid fragment encoding a putative protective antigen against Mycoplasma hyopneumoniae or related infections, said nucleic acid fragment including the following sequence, mutants, derivatives, recombinants and fragments thereof:

30

	10	20	30	40	50	
	1234567890	1234567890	1234567890	1234567890	1234567890	
	ATGAAAAAA	TGCCACTATA	CCAGAGGAAA		TAAAATAATT	50
5	AAAATTACAT	TTTCTTCATT	TGCGCCAGAA	TTTTTAAGAA	TTAGTACATT	100
~	AAAAAGTAGA	ACAAAAGTTA	TTAATGTAAA	CATTAGCGCA	ATCCTTAAGA	150
	AAAATTAAA	AGTITTATCT	ATTTTTTTA	ATCGAAATCC	AACCAGGCAT	200
	AAATCTTTGT	CAGTATTTAT	CAAGTCGGTA	TITTICATT	ATTICTACTA	250
	TTATTATA	TGAATTTGCA	TTTTCCATAA	TCTAAAATTT	TACATTITIT	300
	TATAACAATT	TTAAAAATT	ACTOTITAAT	TTATAGTATT	TITTATITE	350
	TTAGTCTAAA	TTATAAAATT	ATCTTGAATT	TTATTTGAAT	TTTATAATT	400
	TAGTACTAAA	AAATACAAAT	ATTTTTTCCT	ATTCTAAGAA	AAATTCATTT	450
	TTTAAAAAAA	ATTGATTITT	ATAGTATAAT	TIGTITGTAT	AATTGAATTA	500
10	ACTTGATTTG	AAAGGGAACA		AATGCTTAGA	AAAAAATTCT	550
. •	TGTATTCATC	AGCTATTIAT	GCAACTTCGC	TTGCATCAAT	TATTGCATTT	600
	GTTGCAGCAG	GTTGTGGACA		GGTTCAACTT	CTGATTCTAA	650
	ACCACAAGCC	GAGACGCTAA		AAGTAATGAT	TCTATTCGAA	700
	TAGCACTAAC	CGATCCGGAT	AATCCTCGAT	GAATTAGTGC	CCAAAAAGAT	750
	ATTATTICTT	ATGTTGATGA	AACAGAGGCA	GCAACTTCAA	CAATTACAAA	800
	AAACCAGGAT	GCACAAAATA	ACTGACTCAC	TCAGCAAGCT	AATTTAAGCC	850
	CAGCGCCAAA	AGGATTTATT	ATTGCCCCTG	AAAATGGAAG	TGGAGTTGGA	900
	ACTGCTGTTA	ATACAATTGC	TGATAAAGGA	ATTCCGATTG	TTGCCTATGA	950
15	TCGACTAATT	ACTGGATCTG	ATAAATATGA	TIGGTATGTT	TCTTTTGATA	1000
	ATGAAAAAGT	TGGTGAATTA	CAAGGTCTTT	CACTTGCTGC	GGGTCTATTA	1050
	GGAAAAGAAG	ATGGTGCTTT	TGATTCAATT	GATCAAATGA	ATGAATATCT	1100
	AAAATCACAT	ATGCCCCAAG	AGACAATTTC	TTTTTATACA	ATCGCGGGTT	1150
	CCCAAGATGA	TAATAATTCC	CAATATTTTT	ATAATGGTGC	AATGAAAGTA	1200
	CTTAAAGAAT	TAATGAAAAA	TTCGCAAAAT	AAAATAATTG	ATTTATCTCC	1250
	TGAAGGCGAA	AATGCTGTTT	ATGTCCCAGG	ATGAAATTAT	GGAACTGCCG	1300
	GTCAAAGAAT	CCAATCTTTT	CTAACAATTA	ACAAAGATCC	AGCAGGTGGT	1350
	AATAAAATCA	AAGCTGTTGG	TTCAAAACCA	GCTTCTATTT	TCAAAGGATT	1400
20	TCTTGCCCCA	AATGATGGAA	TGGCCGAACA	AGCAATCACC	AAATTAAAAC	1450
	TTGAAGGGTT	TGATACCCAA	AAAATCTTTG	TAACTCGTCA	AGATTATAAT	1500
	GATAAAGCCA	AAACTTTTAT	CAAAGACGGC	GATCAAAATA	TGACAATITA	1550
	TAAACCTGAT	AAAGTTTTAG	GAAAAGTTGC	TGTTGAAGTT	CTTCGGGTTT	1600
	TAATTGCAAA	GAAAAATAAA	GCATCTAGAT	CAGAAGTCGA	AAACGAACTA	1650
	AAAGCAAAAC	TACCAAATAT	TTCATTTAAA	TATGATAATC	AAACATATAA	1700
	AGTACAAGGT	ATATAAAAA	ATACAATTIT	AGTAAGTCCA	GTAATTGTTA	1750
	CAAAAGCTAA	TGTTGATAAT	CCTGATGCCT	AA		1782

54. A method for producing an antibody against a <u>Mycoplasma</u> including providing a biological sample taken a short time after an immune animal has been challenged with a <u>Mycoplasma</u> or <u>Mycoplasma</u> extract taken from the infection site or an area of a lesion or an area close to the infection site or lesion;

isolating cells from the biological sample;

culturing cells <u>in vitro</u> in a suitable culture medium; and harvesting antibodies produced from said cells.

- 55. A method according to claim 54 wherein the biological sample is taken approximately 2 to 7 days after the animal has been challenged with the Mycoplasma.
- 56. A method according to claim 55 wherein the culturing of cells in vitro further includes addition of helper factors to the culture, said helper factors
 selected from the group including cytokines used alone or in combination, including interleukin 1, 2, 3, 4, 5, 6, 7 and 8, colony stimulating factors, interferons and any other factors that may be shown to have an enhancing effect on specific B cell secretion.
- 57. A method according to claim 56 further including a cell activation step including activating the cells isolated to proliferate and secrete and/or release antibodies, said cell activation step including adding a cell activating agent to the culture medium, said cell activating agent selected from the group including mitogens.
 and helper factors produced by leukocytes, or their synthetic equivalents or combinations thereof.
 - 58. A method according to claim 57 wherein the antibody is in the form of the supernatant harvested from the culture medium.
- 25 59. An antibody against a <u>Mycoplasma</u> prepared according to the method of claim 54.
 - 60. A method of identifying a putative protective antigen associated with a Mycoplasma, said method including
- 30 providing

a sample of a Mycoplasma; and an antibody probe including at least one antibody against a

Mycopiasma;

probing the <u>Mycoplasma</u> sample with the antibody probe to detect at least one antigen; and

isolating the antigen detected.

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61. A method of purifying a putative protective antigen associated with a <u>Mycoplasma</u>, said method including providing

a crude antigen mixture; and

an antibody against a Mycoplasma immobilized on a suitable

10 support;

subjecting the crude antigen mixture to affinity chromatography utilizing the immobilized antibody; and

isolating the purified antigen so formed.

15 62. A method for preparing a synthetic antigenic polypeptide against Mycoplasma, which method includes

providing

a cDNA library or genomic library derived from a sample of Mycoplasma; and

an antibody probe including an antibody prepared according to claim 54;

generating synthetic polypeptides from the cDNA library or genomic library:

probing the synthetic polypeptides with the antibody probe; and isolating the synthetic antigenic polypeptide detected thereby.

63. A method according to claim 62 wherein the antibody probe includes an antibody raised against an antigen against <u>Mycoplasma hyopneumoniae</u>, or related infections selected from the group of antigens having approximate molecular weights of 110-114, 90-94, 72-75, 60-64, 52-54 and 46-48 kilodaltons (kD), mutants, derivatives and fragments thereof.

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- 64. A synthetic putative protective antigen produced by the method of claim 62.
- 65. A vaccine or veterinary composition including a prophylactically effective amount of at least one putative protective antigen against a Mycoplasma according to claim 45.
 - 66. A vaccine or veterinary composition according to claim 65 including a plurality of putative protective antigens.
 - 67. A vaccine or veterinary composition including an antibody against a Mycoplasma according to claim 59.
 - 68. A diagnostic kit including an antigen according to claim 45.
 - 69. A method for preventing or treating a <u>Mycoplasma</u> infection, which method including administering to an animal a prophylactically or therapeutically effective amount of at least one putative protective antigen according to claim 45.
 - 70. An isolated DNA fragment encoding a putative protective antigen against <u>Mycoplasma</u> or related infections, said DNA fragment having a nucleic acid sequence according to Figure 6 or an homologous sequence, and functionally active fragments, mutants, variants or recombinants thereof.
 - 71. A clone including a DNA fragment according to claim 70.
 - 72. A clone according to claim 71 which is clone pC1-2
 - 73. An amino acid sequence or functional equivalent thereof encoded by the DNA fragment according to claim 70.

74. An amino acid sequence or functional equivalent thereof having an amino acid sequence according to Figure 7.

REMARKS

Claims 1 - 43 have been cancelled and replaced with new claims 44 - 74. The recitations in claims 44 - 74 correspond to recitations in the claims originally filed as shown on the marked-up copy of the original claims annexed hereto. Accordingly, the new claims draw clear support from the specification as filed.

An early examination of the application is now respectfully requested.

Respectfully submitted,

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ANTIGEN COMPOSITION AGAINST MYCOPLAS

The present invention relates to protective and diagnostic antigens, the preparation thereof, and their use in the formation of vaccine compositions. particularly vaccine compositions against Mycoplasma invodneumoniae infections.

Mycoplasma hyppheniae is a upiquitous swine respiratory pathogen causing mycoplasmal pneumoniae in swine (swine enzootic pneumonia). Swine enzootic pneumonia is probably the most widespread and economically significant disease in swine producing countries of the world. The economic effects of swine enzootic pneumonia (SEP) are complex, and the cost of the disease is severe. In Australia, the disease was estimated in 1988 to cost approximately \$20,000,000 per annum. Increased mortality, decreased crowth weight, depressed feed conversion, susceptibility to secondary bacterial infections, increased management costs, and increased use of antibiotics, are the main reasons for the economic impact of SEP.

Whilst several experimental vaccines have been produced, these have resulted in less than optimal results, and utilising various classes of antibiotics such as tetracycline, lineamycin and tiamulin is still the most widespread control treatment. Such antibiotics are, however of limited therapeutic value, because they do not prevent the establishment of an infection, and lung lesions may develop after treatment ends.

European Patent Application 359,919 to ML Technology Ventures L.P. describes a series of antigens, 36 kD, 41 kD, 74.5 kD and 95 kD in size, and proposes the use of such antigens in vaccines. Results presented suggest that some protection in pigs against challenge was achieved.

However, there remains a need in the art for an effective vaccine against M. hvooneumoniae which would confer protection against colonisation and clinical disease following M, hyponeumoniae challenge and also significantly reduce the morpidity and monality from secondary infections.

Accordingly, it is an object of the present invention to overcome, or at least alleviate, one or more of the difficulties and deficiencies in the prior art

Accordingly, in a first aspect of the present invention there is provided a putative protective antigen against a Mycoplasma, prefetably Mycoplasma

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<u>hyopneumonize</u> prepared by a method including providing

a sample of a Mycoplasma;

an antibody probe including at least one antibody against a Mvcoplasma produced by a method including;

providing a biological sample taken a short time after an immune animal has been challenged with a <u>Mvcoplasma</u> or <u>Mvcoplasma</u> extract taken from the infection site or an area of a lesion or an area close to the infection site or lesion:

10 isolating cells from the biological sample;

culturing cells in vitro in a suitable culture medium; and harvesting antibodies produced from said cells;

probing the <u>Mycoplasma</u> sample with the antibody probe to detect at least one antigen; and

isolating the antigen detected.

The protective antigens may also function as diagnostic antigens as discussed below.

Accordingly, in a preferred aspect of the present invention there is provided a putative protective antigen against <u>Mycoplasma hypopheumoniae</u>, or related infections, selected from the group of antigens having approximate molecular weights of 110-114, 90-94, 72-75, 60-64, 52-54 and 46-48 kilodaltons (kD), as hereinafter described, mutants, derivatives and fragments thereof. The putative protective antigen may be a surface protein. The putative protective antigen may be a surface protein.

25 Preferably the protective antigens are selected from the group of antigens having approximate molecular weights of 110-114, 90-94, 74, 62, 52 and 48 kD.

Preferably, the 72-75 kD antigen includes the following N-terminal amino acid sequence:

AGXLQKNSLLEEVWYLAL

30 and, optionally, one or more of the following internal amino acid sequences:

AKNFDFAPSIQGYKKIAHEL

NEKPEQILOLLG

LLKAEXNKXIEEINTXLDN

Preferably, the 60-64 kD antigen includes one of the following N-terminal amino acid sequences:

MKLAKLLKGFX(N/L)(M/V)IK

5 ADP(F/I)(R/E)Y(V/A)PQG(Q/A)X(M/N)VG

Preferably, the 52-54 kD antigen includes the following N-terminal amino acid sequence;

AGXWAKETTKEEKS

and, optionally, one or more of the following internal amino acid sequences:

10 AWVTADGTVN

AIVTADGTVNDNKPNQWVRKY.

Preferably, the 46-48 kD antigen includes the following N-terminal amino acid sequence;

- AGXGQTESGSTSDSKPQAETLKHKV

and, optionally, one or more of the following internal amino acid sequences: 15

TIYKPDKVLGKVAVEVLRVLIAKKNKASR

AEQAITKLKLEGFDTQ

KNSQNKIIDLSPEG

The 46-48 kD antigen may be encoded by a nucleic acid fragment:

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10	20	30	40	50	
1234567890	1234567890	1234567890	1234567890	1234567890	
ATGAAAAAA	TGCCACTATA	CCAGAGGAAA	GAGCAGTATA	TAAAATAATT	50
AAAATTACAT	TTTCTTCATT	TGCGCCAGAA	TTTTTAAGAA	TTAGTACATT	100
AAAAAGTAGA	ACAAAAGTTA	TTAATGTAAA	CATTAGCGCA	ATCCTTAAGA	150
AAAAATTAAA	AGTTTTATCT	ATTTTTTTA	ATCGAAATCC	AACCAGGCAT	200
AAATCTTTGT	CAGTATTTAT	CAAGTCGGTA	TTTTTCATT	ATTTCTACTA	250
TTATTATAAA	TGAATTTGCA	TTTTCCATAA	TCTAAAATTT	TACATTTTT	300
TATAACAATT	TTAAAAATT	ACTOTTTAAT	TTATAGTATT	TTTATTT	350
TTAGTCTAAA	TTATAAAATT	ATCTTGAATT	TTATTTGAAT	TTTATAATT	400
TAGTACTAAA	AAATACAAAT	ATTITICCT	ATTCTAAGAA	AAATTCATTT	450
TTTAAAAAAA	ATTGATTITT	ATAGTATAAT	TTGTTTGTAT	AATTGAATTA	500
ACTTGATTTG	AAAGGGAACA	AAATGAAAAA	AATGCTTAGA	AAAAAATTCT	550
TGTATTCATC	AGCTATTTAT	GCAACTTCGC	TTGCATCAAT	TATTGCATTT	500
GTTGCAGCAG	GTTGTGGACA	GACAGAATCA	GGTTCAACTT	CTGATTCTAA	550
ACCACAAGCC	GAGACGCTAA	AACATAAAGT	AAGTAATGAT	TCTATTCGAA	700
TAGCACTAAC	CGATCCGGAT	AATCCTCGAT	GAATTAGTGC	CCAAAAAGAT	750
ATTATTTCTT	ATGTTGATGA	AACAGAGGCA	GCAACTTCAA	CAATTACAAA	COE
AAACCAGGAT	GCACAAAATA	ACTGACTCAC	TCAGCAAGCT	AATTTAAGCC	250
CAGCGCCAAA	AGGATTTATT	ATTGCCCCTG	****************	TGGAGTTGGA	200

	ACTECTETTA	ATACAATTGC	TGATAAAGGA	ATTCCGATTG	TTGCCTATGA	950
	TCGACTAATT	ACTGGATCTG	A T AAATATGA	TTGGTATGTT	TCTTTTGATA	1000
	ATGAAAAAGT	TGGTGAATTA	CAAGGTCTTT	CACTTGCTGC	GGGTCTATTA	1050
	GGAAAAGAAG	ATGGTGCTTT	TGATTCAATT	GATCAAATGA	ATGAATATCT	1100
5	AAAATCACAT	ATGCCCCAAG	AGACAATTTC	TTTTTATACA	ATCGCGGGTT	1150
	CCCAAGATGA	TAATAATTCC	CAATATTTTT	ATAATGGTGC	AATGAAAGTA	1200
	CTTAAAGAAT	TAATGAAAAA	TTCGCAAAAT	AAAATAATTG	ATTTATCTCC	1250
	TGAAGGCGAA	AATGCTGTTT	ATGTCCCAGG	ATGAAATTAT	GGAACTGCCG	1300
	GTCAAAGAAT	CCAATCTTTT	CTAACAATTA	ACAAAGATCC	AGCAGGTGGT	1350
10	AATAAAATCA	AAGCTGTTGG	TTCAAAACCA	GCTTCTATFT	TCAAAGGATT	1400
	TCTTGCCCCA	AATGATGGAA	TGGCCGAACA	AGCAATCACC	AAATTAAAAC	1450
	TTGAAGGGTT	TGATACCCAA	AAAATCTTTG	TAACTCGTCA	AGATTATAAT	1500
	GATAAAGCCA	AAACTITTAT	CAAAGACGGC	GATCAAAATA	TGACAATTTA	1550
	TAAACCTGAT	AAAGTTTTAG	GAAAAGTTGC	TGTTGAAGTT	CTTCGGGTTT	1500
15	TAATTGCAAA	GAAAAATAAA	GCATCTAGAT	CAGAAGTCGA	AAACGAACTA	1550
	AAAGCAAAAC	TACCAAATAT	TTCATTTAAA	TATGATAATC	AAACATATAA	1700
	AGTACAAGGT	ATATAAAAA	ATACAATTIT	AGTAAGTCCA	GTAATTGTTA	1750
	CAAAAGCTAA	TGTTGATAAT	CCTGATGCCT	AA		1732

Accordingly, in a further aspect the present invention provides an isolated nucleic acid fragment encoding a putative protective antigen against <u>Mycoplasma</u> <u>hyopneumonize</u> or related infections, said nucleic acid fragment:

	10	20	30	40	50	
25	1234557890	1234567890	1234557890	1234557590	1234567890	
			-			
	atgaaaaaa	TGCCACTATA	CCAGAGGAAA	GAGCAGTATA	TAAAATAATT	50
	AAAATTACAT	TTTCTTCATT	TGCGCCAGAA	TTTTTAAGAA	TTAGTACATT	100
	AAAAAGTAGA	ACAAAAGTTA	TTAATGTAAA	CATTAGCGCA	ATCCTTAAGA	150
3C	A÷ATTA÷A	AGTTTTATCT	ATTITITA	ATCGAAATCC	AACCAGGCAT	200
	AGATOTTTGT	CAGTATTTAT	CAAGTCGGTA	TTTTTCATT	ATTICTACTA	250
	ASSTATTATT	TGAATTTGCA	TTTTCCATAA	TOTAAAATTT	TACATTITIT	300
	TATAKCAATT	TTAAAAATT	ACTCTTTAAT	TTATAGTATT	TITIATITI	350
	TTAGTCTAAA	TTATAAAATT	ATCTTGAATT	TTATTTGAAT	TTATAATT	400
35	TAGTACTAAA	AAATACAAAT	ATTTTTCCT	ATTOTAAGAA	AAATTCATTT	450
	TTTAAAAAAA	ATTGATTTTT	ATAGTATAAT	TTGTTTGTAT	AATTGAATTA	500
	ACTTGATTTG	AAAGGGAACA	AAATGAAAAA	AATGCTTAGA	AAAAAATTCT	550
	TGTATTCATC	AGCTATTTAT	GCAACTTCGC	TTGCATCAAT	TATTGCATTT	500
	GTTGCAGCAG	GTTGTGGACA	GACAGAATCA	GGTTCAACTT	CTGATTCTAA	550
40	ACCACAAGCC	GAGACGCTAA	AACATAAAGT	AAGTAATGAT	TCTATTCGAA	700
	TAGCACTAAC	CGATCCGGAT	AATCCTCGAT	GAATTAGTGC	CCAAAAAGAT	750
	ATTATTOTT	ATGTTGATGA	AACAGAGGCA	GCAACTTCAA	CAATTACAAA	006
	AAAECAGGAT	GCACAAAATA	ACTGACTCAC	TCAGCAAGCT	AATTTAAGCC	350
	CAGCGCCAAA	AGGATTTATT	ATTGCCCCTG	AAAATGGAAG	TGGAGTTGGA	900
45	ACTGCTGTTA	ATACAATTGC	TGATAAAGGA	ATTCCGATTG	TTGCCTATGA	9 50
	TEGACTAATT	ACTGGATCTG	ATAAATATGA	TTGGTATGTT	TETTTIGATA	1000
	atgaaaaagt	TGGTGAATTA	CAAGGTCTTT	CACTTGCTGC	GGGTCTATTA	1050
	GGA÷AAGAAG	ATGGTGCTTT	TGATTCAATT	GATCAAATGA	ATGAATATCT	100 ר
	AAAATCACAT	ATGCCCCAAG	AGACAATTTC	TITTATASA	ATCGCGGGTT	1150
50	CCCAAGATGA	TAATAATTCC	CAATATITIT	ATAATGGTGC	AAT GAAAGT A	1200
	CTTAAAGAAT	TAATGAAAAA	TTCGCAAAAT	AAAATAATTG	ATTTATCTCC	1250
	TGAAGGCGAA	AATGCTGTTT	ATGTCCCAGG	ATGAAATTAT	GGAACTGCCG	1300
	GTC÷AAGAAT	CCAATCTTT	CTAACAATTA	ACAAAGATCC	AGCAGGTGGT	1350

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AATAAAATCA AAGCTGTTGG TTCAAAACCA TCTTGCCCCA AATGATGGAA TGGCCGAAC TTGAAGGGTT TGATACCCAA AAAATCTTTG GATAAAGCCA AAACTTTTAT CAAAGACGG TAAACCTGAT AAAGTTTTAG GAAAAGTTG TAATTGCAAA GAAAAATAAA GCATCTAGAA AAAGCAAAAC TACCAAATAT TTCATTTAAA AGTACAAGGT AAAAATATTA ATACAATTTT CAAAAGCTAA TGTTGATAAT CCTGATGCC	A AGCAATCACC AAATTAAAAC 14: TAACTCGTCA AGATTATAAT 15: C GATCAAAATA TGACAATTTA 15: C TGTTGAAGTT CTTCGGGTTT 15: CAGAAGTCGA AAACGAACTA 15: TATGATAATC AAACATATAA 17: AGTAAGTCCA GTAATTGTTA 17:	50 50 50 50 50
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As cross protection between various Mycoplasma such as M. hvorhinis and M. synoviae has been documented, similar antigens may also be detected in other Mycopiasma species (Figure 1).

In a still further aspect the present invention provides a method for preventing Mycoplasma infection in animals. Preferably the Mycoplasma disease is a Mycoplasma hyppneumoniae disease such as swine enzootic pneumonia (SEP). This method includes administering to an animal an effective amount of at least one protective antigen against Mycoplasma as described above.

The present invention further provides a vaccine composition including a prophylactically effective amount of at least one putative protective antigen against a Mycoplasma as herein described. Preferably the veterinary composition includes two or more putative protective antigens as herein described.

Accordingly in a preferred aspect the present invention provides a vaccine composition including two or more putative protective antigens selected from antigens having approximate molecular weights of 110-114, 90-94, 72-75, 60-64, 52-54 and 46-48 kilodattons.

The vaccine composition may include any combination of two or more putative protective antigens selected from antigens having approximate molecular weights of 110-114, 90-94, 72-75, 60-64, 52-54 and 46-48 kD. The two or more antigens may be selected from antigens falling within one of the specified approximate molecular weights and/or antigens from different specified approximate molecular weights. The composition may contain 3, 4, 5 or 6 antigens selected from protective antigens having molecular weights of approximately 110-114, 90-94, 72-75, 50-64, 52-54 and 46-48 kD.

The vaccine compositions according to the present invention may be

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administered orally or may be administered parenterally (for example by intramuscular, subcutaneous, intradermal or intravenous injection). The amount required will vary with the antigenicity of the active ingredient and need only be an amount sufficient to induce an immune response typical of existing vaccines.

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Reactive experimentation will easily establish the required amount. Typical initial doses of vaccine or veterinary compositions may be approximately 0,001-1 mg active ingredient/kg body weight. The dose rate may increase or multiple doses may be used as needed to provide the desired level of protection.

The vaccine composition according to the present invention may further include a veterinary acceptable carrier, diluent or excipient therefor. Preferably the active ingredient may be suspended or dissolved in a carrier. The carrier may be any solid or solvent that is nontoxic to the animal and compatible with the active ingredient. Suitable carriers include liquid carriers, such as normal saline and other nontoxic salts at or near phsylological concentrations, and solid carriers, such as talc or sucrose.

Preferably the vaccine contains an adjuvant, such as Freund's adjuvant, complete or incomplete, or immunomodulators such as cytokines may be added to enhance the antigenicity of the antigen if desired.

More preferably the adjuvant is of the mineral-oil type as these have been found to be consistently superior at inducing antibody titres and Delayed Type Hypersensitivity responses. A particularly preferred adjuvant is that marketed under the trade designation Montanide ISA-50 and available from Seppic, Paris, France.

When used for administering via the pronchial tubes, the vaccine is suitably present in the form of an aerosol.

In a still further aspect of the present invention there is provided a diagnostic kit including a diagnostic antigen against a <u>Mycoplasma</u>, preferably <u>Mycoplasma</u> hypneumoniae, identified and purified as described above.

The putative protective antigens according to the present invention may be isolated and identified utilising the general methods described in Australian patent application 49035/90, the entire disclosure of which is incorporated herein by reference

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Accordingly, in a further aspect, the present invention provides a method for producing at least one antipody against a <u>Mycoplasma</u>. This method includes providing a biological sample taken a short time after an immune animal

has been challenged with a <u>Mycoplasma</u> or <u>Mycoplasma</u> extract taken from the infection site or an area of a lesion or an area close to the infection site or lesion;

isolating cells from the biological sample;

culturing cells <u>in vitro</u> in a suitable culture medium; and harvesting antibodies produced from said cells.

The Mycoplasma may be Mycoplasma hyooneumoniae.

The animal may be a mammal including humans. The mammal may be a comestic animal such as a pig, sheep or cattle.

The biological animal sample may be of any suitable type. The biological sample may be taken from animal tissue, organs, lymph or lymph nodes. The biological sample may be taken from the infection site, the lungs of the animal, or an area of a lesion which may be formed or an area close to the infected site or a lesion such as in the lymph nodes draining from the lungs.

However, serum/plasma samples are not used as the biological samples according to this aspect of the present invention. It has been found that the majority of antibodies found in a serum/plasma sample are irrelevant to protection or specific diagnosis or a <u>Mycoplasma</u> or are unrelated to the <u>Mycoplasma</u>. In addition, other serum/ plasma components may interfere with the specific reactions between pathogen components and antibodies to them.

In contrast, the probes described in the present invention are highly enriched in <u>Mycoplasma</u>-specific antibodies of particular importance to protective immunity.

It is preferred that the biological samples are taken from the animals at a predetermined time in the development of the disease. In general, for a <u>Mycoplasma</u> infection, it has been found that the biological samples should be taken approximately 2 to 7 days after challenge with or after administration of products obtained from a pathogen or with the pathogen itself.

The cells isolated from the biological sample may include B cells.

Thus, preferably the cells are taken a short time after in vivo stimulation.

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preferably within approximately 2 to 5 days thereafter, resulting in the <u>in vivo</u> induction of antibody forming cells which will secrete specific antibodies into the culture medium after <u>in vitro</u> incubation.

In vitro secretion of antibodies in the culture medium by recently activated B cells may be enhanced by the addition of helper factors to the cultures. The helper factors may be cytokines used alone or in combination, including Interleukin 1, 2, 3, 4, 5, 6, 7 and 8, colony stimulating factors, interferons and any other factors that may be shown to have an enhancing effect on specific B cell secretion.

The method of producing an antibody may include a further step of activating the calls isolated to proliferate and secrete and/or release antibodies.

The cell activation step may include adding a cell activating agent to the culture medium. The cell activating agent may be selected from mitogens and helper factors produced by leukocytes, or their synthetic equivalents or combinations thereof.

The mitogens may be selected from the group including products derived from pokeweed (Phytolacca americana) also known as pokeweed mitogen (PWM), polyvinylpyrrolidone (PVP), polyadenylic-polyuridylic acid (poly(A-U)), purified protein derivate (PPD), polyinosinic-polycytidilic acid (poly(I-C)). lipopolysaccharide (LPS), staphylococcal organisms or products thereof. Bactostreptolysin O reagent (SLO), Staphylococcal phage lysate (SPL), Epstein-Barr virus (EBV), Nocardia water-soluble mitogen (NWEM), phytohemagglutinin (PHA). Concanavalin A (Con A), and dextran-sulphate and mixtures thereof. The cell proliferation agent may be any agent that indirectly or directly results in B cell proliferation and/or antibody secretion such as solid-phase anti-immunoglobulin. The helper factors may be selected from the group including cytokines including interleukin 1, 2, 3, 4, 5, 6, 7 and 8, colony stimulating factors, interferons and any other helper factors that may be shown when added alone, or in combination with other factors and agents, to have an enhancing effect on specific B cellproliferation and/or antibody secretion. This in no way is meant to be an exhaustive list of mitogens and cell actuating agents including helper factors.

The in visto culturing of the cells may be concusted with or without prior

steps to separate sub-populations of cells. The harvesting of antibodies may be conducted by harvesting of the supernatant from the culture medium. This supernatant contains antibodies secreted by these cells during the in vitto culture or artificially released from the B cells, for example by lysis of the B cells. It has been found that the antibody-containing supernatants may be used directly to detect antigens of the Mycoplesma.

in a preferred aspect of the present invention, there is provided a method for identifying an antigen associated with a <u>Mycoplasma</u>, preferably <u>Mycoplasma</u> <u>hyopneumoniae</u>. This method includes

10 providing

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a sample of a Mycoplasma; and

an antibody probe including at least one antibody against a Mycoplasma;

probing the <u>Mvcoplasma</u> sample with the antibody probe to detect at least one antigen; and

isolating the antigen detected.

The sample of <u>Mycoplasma</u> may be mixed with a standard buffer solution and placed on a standard support such as an SDS-polyacrylamide gel to separate the proteins contained thereon (Figure 2)

Alternatively the proteins may be selected utilising the non-ionic detergent Triton X-114 (TX-114). Insoluble material may be removed by centrifugation. Proteins soluble in the TX-114 phase may then be precipitated out (Figure 2).

The separate proteins may then be transferred to nitrocellulose, nylon or other sheets.

The probing with a suitable antibody may further include subjecting the product produced thereby to a detection assay. The detection assay may include Western plot techniques. The detection assay may be an immunoprecipitation assay a radioimmunoassay, an enzyme-linked immunoassay or immunofluorescent assay (Figures 3, 4 and 5).

The antibody produced as described above may be utilized simply in the form of the supermatant harvested from the culture medium. Alternatively, the antibodies may be separated and putified

- 10 -

in a further preferred espect of the present invention the antibody contained in the culture medium may be used for the affinity purification, preferably immuno-affinity purification of antiqen.

Accordingly, in a preferred aspect there is provided a method for purifying antigen. This method includes

providing

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a crude antigen mixture; and

an antibody against a <u>Mycoplasma</u> immobilized on a suitable support;

subjecting the crude antigen mixture to affinity chromatography utilizing the immobilized antibody; and

isolating the purified antigen so formed.

The antibody is produced by the method described above.

Antibody can be obtained from the culture supernatant probe by conventional methods. For example, methods usually used to purify immunoglobulins from serum or plasma, e.g. precipitation with ammonium sulphate, fractionation with caprylic acid, ion exchange chromatography, or by binding and elution from immobilized protein G or protein A, may be utilized. Antibody so obtained can then be coupled to suitable supports, e.g., CNBractivated Sepharose 43 (Pharmacia), Affi-gel (Bio-RAD), or other affinity chromatography supports able to bind proteins.

Immobilized antibody can then be applied to the fractionation and purification of specific antigen from a complex <u>Mycoplasma</u> extract by affinity chromatography. After binding of antigen to immobilized antibody, unbound macromolecular species can be washed away from the solid support with, e.g. buffers containing 1.5 M NaCl. Subsequently the antigen can be eluted from the affinity column with, e.g. low or nigh pH buffer or buffers containing chaotropic ions, e.g. 0.5-3.0 M sodium thiocyanate.

The application of the antibody probe to affinity chromatography enables sufficient quantities of specific antigens to be rapidly isolated from a complex crude extraction mixture for biochemical characterization, amino-acid sequencing and vaccination of animal for limited protection studies. Application of affinity

chromatography for obtaining antigen(s) avoids the difficulties often encountered when applying conventional biochemical techniques to the purification of an antigen about which little or no data is known. It also obviates the need to raise polyclonal or monoclonal antibodies for the purpose of "analytical" affinity chromatography. Large scale preparation may, however, require the preparation of polyclonal or monoclonal antibodies.

Having identified the antigen(s) molecular biology, chemical techniques, e.g. cloning techniques, may be used to produce unlimited amounts of this antigen or, alternatively, synthetic peptides corresponding to different fragments of the identified antigens may be used as a means to produce a vaccine.

Accordingly in a preferred aspect of the present invention there is provided a method for preparing a synthetic antigenic polypeptide against <u>Mycoplasma</u>, preferably <u>Mycoplasma</u> hypneumoniae, which method includes

providing

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a cDNA library or genomic library derived from a sample of Mycoplasma; and

an antibody probe as described above;

generating synthetic polypeptides from the cDNA library or genomic library; probing the synthetic polypeptides with the antibody probe; and

isolating the synthetic antigenic polypeptide detected thereby.

Either cDNA or genomic libraries may be used. The cDNA or genomic libraries may be assembled into suitable expression vectors that will enable transcription and the subsequent expression of the clone cDNA, either in prokaryotic hosts (e.g. bacteria) or eukaryotic hosts (e.g. mammalian cells). The probes may preferably be selected from

- (i) synthetic oligonucleotide probes based on the amino acid sequence of the antigen identified and punified as described above.
- (ii) antibodies obtained from the culture medium produced as described above:
- 30 (iii) monoclonal or polyclonal antibodies produced against the antigens identified and purified as described above;
 - (w) recombined; or synthetic monoclonal antipodies or polypeptides with

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specificity for the entigen, e.g. as described by Ward et al., Nature, 241, pages 544-546 (1989).

The synthetic antigenic polypeptide produces in accordance with the invention may be a fusion protein containing the synthetic antigenic peptide and another protein.

In a further aspect of the present invention there is provided a DNA fragment encoding a putative protective antigen against Mycoplasma or related infections, said DNA fragments having a nucleic acid sequence according to Figure 6a and 6b or an homologous sequence and functionally active fragments thereof.

In a further preferred aspect of the present invention there is provided a cione including a DNA fragment encoding a putative protective antigen against Mycoplasma or related infections, said DNA fragments having a nucleic acid sequence according to Figure 6a and 6b or an homologous sequence and functionally active fragments thereof.

Preferably the clone is oC1-2.

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The present invention will now be more fully described with reference to the accompanying Examples and drawings. It should be understood, however, that the description following is illustrative only and should not be taken in any way as a restriction on the generality of the invention described above.

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IN THE FIGURES:

FIGURE 1: SDS-Polyacrylamide gel (12.5%) profiles of SDS extracts of species of mycoplesma- Coomassie R250 stained.

30

i ensi Pre-stained Molecular Weight Standards

M. gallisepticum Lane 2

Lane 3 M. synoviae.

Lane 4 M. hyopneumoniae.

Lane 5 M. hyorhinis.

Lane 6 M. flocculare.

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FIGURE 2: SDS-Polyacrylamide gel (12.5%) profiles of extracts of strains of M. hyppneumonize - Coomassie R250 stained gel

- 13 **-**

Lane 1 Pre-stained Molecular Weight Standards.

10 Lane 2 Triton X-114 extract of M. hyppneumoniae - strain Beaufort.

Lane 3 As for Lane 2.

Lane 4 SDS extract of M. hyopneumoniae strain Beauton.

Lane 5 SDS extract of M. hyopneumoniae strain 10110.

15 <u>FIGURE 3</u>: Western blots of Triton X-114 extracted antigens from M. hyppneumoniae strain Beaufort, probed with serum and supermatant antibody probes.

Lane 1 No antibody control

20 Lane 2 Dookie pig serum control 1/200.

Lane 3 Pig 105 supernatant.

Lane 4 Pig 1 supernatant.

Lane 5 Dookie pig supernatant.

25 <u>FIGURE 4</u>: Western blots of SDS extracted antigens from *M. hyopneumoniae* strain Beaufort probed with paired serum and supernatantantibody probes. Fractionation of antigens on SDS Polyacrylamide gel (12.5%).

Lane 1 a) Pig 453 supernatant.

30 b) Pig 453 serum 1/100.

Lane 2 a) Pig 105 supermanant.

b) Prg 105 serum 1/100.

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Lane 3	a)	Pig -	l superanatant.

b) Pig 1 serum 1/100.

Lane 4 a) Pig 15 supermanant.

b) Pig 15 serum 1/100.

5 Lane 5 a) Dookie supernatant.

b) Dookie serum 1/100.

Lane 6 No antibody control.

FIGURE 5: Western blots of SDS extracted antigens from *M. hyopneumoniae* strain Beaufort probed with paired serum and supernatantantibody probes. Fractionation of antigens on SDS Polyacrylamide gel (10.0 %).

Lane 1 a) Pig 453 supernatant.

b) Pig 453 serum 1/100.

15 Lane 2 a) Pig 105 supermatant.

b) Pig 105 serum 1/100.

Lane 3 a) Pig 1 supematant.

o) Pig 1 serum 1/100.

Lane 4 a) Pig 15 supernatant.

o) Pig 15 serum 1/100.

Lane 5 a) Dookie supernatant.

b) Dookie serum 1/100.

Lane 6 No antibody control.

25 FIGURE 6: The entire 48 k gene sequence.

FIGURE 7: tHE 48kDa protein sequence of the 48k gene sequence.

PCT/AT196/0014

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EXAMPLE 1

Mycoplasma hyopneumoniae media

Friss Media

Hovind-Hougen, K., Friss, N.F., Research in Veterinary Science, 1991, 51, pp. 155-163, "Morphological & Ultrastructural Studies of M. flocculare and M. hyppneumoniae in vitro".

250 mi. Hanks BSS

10 140 m! Water

1.5 gm Brain Heart infusion

1.6 gm PPLO Broth w/o CV

Autociave at 120°C for 20 minutes

18 ml Yeast Extract (100g YSC-2 Sigma in 750 ml)

15 3.7 ml 0.2% DNA in 0.1% Na₂CL₃

5.14 ml 1% -NAD

0.6 ml 1% Phenoi red

Adjust to pH 7.3 to 7.4

20 Filter through 0.45 um, 0.2 um membrane, store at 4°C.
Add sterile Horse or Pig serum to 20%
and Antibiotics prior to use

Etheridae Media

Etheridge, J.R., Cottew, G.S., Lloyd, L.C., Australian Veterinary Journal, 1979. August 55, pp 356-359, "Isolation of Mycoplasma hypopheumoniae from lesions in experimentally infected pigs".

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	<u>Materials</u>	For 600 mls
	Hanks BSS	18.9 m!
	Hartleys Digest broth	1.28 gm
5	Heart Infusion broth	1. 35 gm
	Lactalbumin hydrolysate	2.21 gm
	Glucose	4.41 gm
	Yeast Extract autolysate	8.82 ml
	Pig Serum (filtered)	163 ml
10	1% NAD	6.17 ml
	1% Phenol red	1.32 ml
	0.2% DNA in 0.1% Na ₂ CO ₃	4.41 ml

Make up to 600 ml with MQ water (about 350 - 400 ml)

Adjust pH to 7.4 and fifter through: 3.0 um, 0.8 um 0.45 um, 0.2 um. Store at 4°C.

Development of Immune Sows

Cull sows and naive gilt (unmated sow designated Dookie).

20 Challenged on numerous occasions, with culture grown M. https://hypneumoniae.com/hypneumoni

Tiamulin antibiotic given 31st January, 1992 to 4th February, 1992. Rested for approximately 8 weeks.

25 Infectious Challenge

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120 ml of frozen culture of <u>M. hyppheumonize</u> strain Beaufort spun down (12,000 xg. 20 min.) and resuspended in 50 ml complete medium and cultured overnight at 37°C. The overnight culture was centrifuged (12,000 xg. 20 min.) and the <u>Mycoplasma</u> cells resuspended in 10 ml serum free <u>Mycoplasma</u> culture medium. The 10 ml of concentrated mycoplasma was administered to anaesthetised immune sows via a catheter to ensure the inoculum was placed into the trappes.

Three of four days post-challenge, the sows were killed, and lymph nodes draining the lungs taken - these included the left and right tracheobronchial lymph nodes, and the lymph nodes located at the bifurcation of the trachea.

Antibody probes were prepared from pig lymph nodes and utilised to detect putative protection antigens as described in Australian Patent Application 49035/90 referred to above. Separate cell cultures were obtained from individual lymph nodes. Culture supernatants were harvested after 5 days of culture.

Antioen Preparation

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Mycoplasma hyopneumoniae strain Beaufort was cultured in Etheridge media until the pH had dropped to between 6.8 and 7.0. Cells of M. hyopneumoniae were harvested from culture by centrifugation at 12,000 xg for 20 min., washed 4 times with either sterile PBS or 0.25 M NaCl and then the pelleted cells extracted with one of the following.

(i) Socium dodecvi sulphate (SDS)

The cell pellet was resuspended in 0.2% SDS and extracted for 2 hours at 37°C. Insoluble material was pelleted from the extract at 12,000 xg for 10 min. and the soluble extract run on SDS-polyacrylamide gel electrophoresis (SDS-PAGE).

(ii) <u>Triton X-114</u>

The method of Bordier (J. Bio, Chem. 1981, 255:1604-1606) was used to selectively extract memorane proteins using the non-ionic detergent Triton X-114.

The cell pellet was resuspended in cold PBS to 2 mg/ml protein and a cold pre-condensed solution of TX-114 added to give a final concentration of 1% (v/v) TX-114. Extraction was achieved by incubation overnight at 4°C with gentle mixing. Insoluble material was removed by centrifugation at 12,000 xg for 20 min. at 4°C. The Triton X-114 soluble membrane proteins were then obtained by achieving a phase separation at 37°C.

Proteins soluble in TX-114 phase were predictated with 80% ethanol in the presence of parrier dexiran (80,000 molecular weight) at -70°C overnight. The proteins were collected by centrifugation at 12,000 kg for 30 min. and dissolved to 500 ug/ml in 4 M urea.

Identification of Anticens

Six antigens were identified utilising the above-mentioned technique. The identified antigens were those that were consistently identified by the antibody probes from the immune cultures and the Dookie gilt. The results are summarised in Table 1.

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		TABLE 1
	Molecular Weight (kD)	Characteristics
	110-114	SDS Extracted
	90-94	SDS Extracted
10	72-75	Triton X-114 Extracted
	50-64**	SDS Extracted. Partitions to aqueous
		phase of Triton X-114 extract.
	52-54	Triton X-114 Extracted
	46-48	Triton X-114 Extracted
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Two antigens of approximate molecular weight 62 kD were identified.

	<u> Molecular</u>	<u>Amino Acid Sequence</u>
	Weight (kD)	
20		
	46-48	48 K N-Terminal: AGXGQTESGSTSDSKPQAETLKHKV
		48 K CNSR F 1: TIYKPDKVLGKVAVEVLRVLIAKKNKASR
		48 K CNBR F 2: AEQAITKLKLEGFDTQ
		48 K CNBR F 3: KNSQNKIIDLSPEG
25		
	52-54	52 K N-Terminal, AGXWAKETTKEEKS
		52 K CNBR F 1. AWVTADGTVN
		52 K CNBR F 2: AIVTADGTVNDNKPNQVVRKY
30	60-64	52 K N-Terminal: MKLAKLLKGFX (N/L)(M/V) IK
	5 0-54	52 K N-Terminal ADP(F/i)(R/E)Y(V/A)PQG(Q/A)X(M/N)VG

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72-75 74 K N-Terminal: AGXLQKNSLLEEVWYLAL

74 K CNBR F 1: AKNFDFAPSIQGYKKIAHEL

74 K CNBR F 2: NLKPEQILQLLG

5 74 K CNBR F 3: LLKAEXNKXIEEINTXLDN

CNBR - Cyanogen Bromide fragment X denotes an undetermined amino acid (A/B) - residue may be A or B

PCR of 48kDa Gene

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Polymerase Chain Reaction (PCR) oligonucleotide primers were designed from the amino acid sequences obtained from the N-terminal and internal cyanogen bromide (CNBr) derived peptides. Inosine (I) was substituted at positions of high redundancy. The following primers were used in a standard PCR assay, run on a Bartelt Gene Machine Robotic thermal cycling instrument.

Oligo 48 K CNBr F 1: ACIAACGACGAGAAGCCICAGGC

TTAAAA

20 Oligo 48 K CNBr F 2: TTIAGCTTIGTGATIGCCTGCTC

AT A T T

-

Oligo 48 K CNBr F 3: AGGTCGATGATCTTCCAICC

25 AA A A T T T T

The resulting PCR products were visualised on a 1.5% agarose gel, excised, and purified using Prep-a-Gene (BioRad). They were cloned by standard techniques into a dideoxy tailed T-vector (Holton and Graham, Nucleic Acids Research 19, 1156, 1991) and the nucleic acid sequence determined. The PCR product, obtained from the reaction using primers F1 and F2 shown above, was of

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approximately 810 base pairs and was shown by sequencing to code for the previously determined amino acid sequence of the purified native 45-48kDa protein.

5 Genomic clone isolation at 48 k dene

The entire 48k gene and 48kDa protein (Figures 6 and 7) has been isolated and sequenced. The gene was obtained from an *M. hyopneumoniae* genomic library made by digesting genomic DNA with the restriction enzyme CLA I and ligating the fragments into the vector pBluescript (Stratagene). The ligated product was then electroporated into *Escherichia coli* strain SURE (Stratagene) and the cells plated on Luria Broth agar plates containing 100 µg/ml Ampicillin (LB-Amp). The library was screened by DNA hybridisation with a polymerase chain reaction (PCR) product specific for the 48 kDa protein. Positive clones were grown in LB-Amp, the cells harvested and the DNA isolated and partially sequenced for confirmation.

The positive clone pC1-2 was entirely sequenced and the protein sequence deduced. This was compared to the protein sequence obtained from the N terminus and Cyanogen Bromide fragments of the 43 kDa protein to show the that the gene encoded the desired protein.

Adjuvant Selection

Young piglets, 5-7 weeks of age, were immunised with identified antigen(s). The antigens include Triton X-114 extract and identified proteins of 46-48, 52-53, 60-64, 70-75, 90-94 and 110-114 kD, either singly or in combination. An immunising dose of antigen, containing between 5-100 µg protein, was given by intramuscular injection in combination with an adjuvant. An adjuvant is selected from

- (i) Seppic Montanise ISA-50
- (ii) Quill A and other derivatives of saponin.
 - (iii) oil in water emulsion employing a mineral oil such as Bayol F/Ariacel A.
 - (iii) oil in water emulsion employing a vegetable oil such as com oil

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safflower oil or other with lecithin as emulsifier,

- (v) aluminium hydroxide gel, and
- (vi) nonionic block polymer such as Piuronic F-127 produced by BASF (U.S.A.).
- Immunising doses were given at 2-4 week intervals, the number of doses being dependent on the adjuvant and amount of antigen, but preferably 2 to 3 doses are given.

Adjuvants were treated on the basis of being able to induce antibody titres, as measured by ELISA, and by assessment of induced cell-mediated immunity as tested by Delayed-Type Hypersensitivity (DTH) reaction.

The results clearly show that <u>mineral-oil type adjuvants are consistently superior</u> at inducing antibody titres and DTH responses (Table 2). In particular an adjuvant marketed under trade designation Montanide ISA-50 and available from Seppic, Paris, France has been found to be suitable.

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TABLE 2

TABLE 2					
GROUP	IsminA	DTH 24 Hour	DTH 48 Hour	Antiboay Levels	
	Number	Response	Response	(450 nm)	
	4,55				
	19	0	0	0.061	
	11	0	0	0.010	
CONTROL	1	-	•	0.005	
(Unvaccinated)	15	0	0	0.038	
	7	О	0	0.005	
			· · · · · · · · · · · · · · · · · · ·		
	18	+	0	0.753	
QUIL A	25	÷	0	0.788	
	17	0	0	0.638	
	158	_	<u></u>	0.642	
	169	÷+÷	0	0.316	
	22	22 0 0		0.521	
VEG. OIL	4	÷	0	0.665	
	5	5 - -		0.239	
	13	÷-÷	÷÷	0. 45 7	
	14	* **	÷÷	1.086	
	5	+++	÷÷	1.024	
MIN. OIL	23	++-	7	0.864	
	15	÷÷÷	O	0.975	
	21	+		0.954	

TABLE 2. Antipody levels and DTH responses in pigs measured 2 weeks after the third injection of antigen from M. hyppneumoniae. (- = no response; _ = faint reddening; + = faint reddening and swelling; -- = reddening; --- = swelling with or without reddening).

Protection Pen Trial

Groups of 9 young piglets, 6 weeks of age, were immunised with purified and semi-putified antigens as shown in Table 3 below. The antigens were purified on reversed-phase HPLC using a formic acid solvent system with an acetonitrile gradient.

Antigens were resolubilised in 4 Molar urea before incorporation in mineral oil adjuvant.

The immunisation schedule is as shown in Table 2.

- 24 **-**

TABLE 3

Protocol for Pen Trial of Antigens of Mycoplasma Hyppneumoniae

5 VACCINATIONS & BLEEDS

Treatment	Day Number
1st Vaccination	0
2nd Vaccination	14
3rd Vaccination	50
Infectious Challenge	64
Slaughter	91

ANTIGEN DOSES

ANTIGEN DOSES	
Partly Purified	1st & 2nd Vacens, 50µg COMPLEX ANTIGEN/DOSE
62 kD	3rd Vaccn 220ug PARTIALLY PURIFIED ANTIGEN/DOSE
(Puritied)74÷52kD	1st Vaccn. 20μg total protein/DOSE
	2nd Vaccn. 13µg total protein/DOSE
	3rd Vacch. 17μg total protein/DOSE
(Purified) 45KD	1st Vaccn. 20µg/DOSE
	2nd Vaccn. 18µg/DOSE

10 ALL PROTEIN ESTIMATIONS DONE BY "BCA" PROTEIN ASSAY (Pierce, Illinois, U.S.A.

3rd Vaccn, 27µg/DOSE

Protection from infection with <u>Mycoplasma hyponeumoniae</u> was assessed by infectious challenge 2 weeks after the final immunisation. Infectious challenge was achieved by intranasal administration of 10ml of a 10% (w/v) lung homogenate, prepared from infected lung, and by housing test piglets with

- 25 -

previously infected piglets. Four weeks after infectious challenge, the animals were killed and the extent and degree of lung lesions assessed (Table 4).

TABLE 4

Pen Trial of Antigens of Mycoplasma Hyopneumonize						
Group No.	No. Pneumonia	Median Lung	% Reduction			
	Free (%)	Lesion Score	(from Median)			
Controls	1 (11)	13	0%			
52 kD	0 (0)	5	61%			
74÷52 kD	3 (33)	6.75	48%			
48 kD	2 (22)	5.25	52%			

REFERENCE

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Warren H.S. and Chedid, L.A., Future Prospects for Vaccine Adjuvants CRC Critical Reviews in Immunology 8: 83-108, 1988.

Finally, it is to be understood that various other modifications and/or alterations may be made without departing from the spirit of the present invention as outlined herein.

CLAIMS:

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A putative protective antigen against a <u>Mycoolasma</u>, prepared by a method
 including

providing

a sample of a Mycoplasma;

an antibody probe including at least one antibody against a Mycoplasma produced by a method including;

providing a biological sample taken a short time after an immune animal has been challenged with a <u>Mycoplasma</u> or <u>Mycoplasma</u> extract taken from the infection site or an area of a lesion or an area close to the infection site or lesion;

isolating cells from the biological sample;

culturing cells in vitro in a suitable culture medium; and harvesting antibodies produced from said cells;

probing the <u>Mycoplasma</u> sample with the antibody probe to detect at least one antigen; and

isolating the antigen detected.

- 2. A putative protective antigen according to claim 1 wherein the Mycoplasma is Mycoplasma hypopheumoniae.
- 3. A putative protective antigen against <u>Mycopiasma hyponeumoniae</u>, or related infections, selected from the group of antigens having approximate molecular weights of 110-114, 90-94, 72-75, 50-64, 52-54 and 46-48 kilodaltons (kD), as herein described, mutants, derivatives and fragments thereof.
- 4 A putative protective antigen according to plaim 3 which is a surface 30 protein.

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- 5. A putative protective antigen according to claim 3 or 4 which is a surface lipo-protein or membrane protein.
- 6 A putative protective antigen according to any one of claims 3-5 having approximate molecular weight of 110-114, 90-94, 74, 62, 52 and 48 kD.
 - 7. A putative protective antigen according to claim 3 wherein the antigen in the 72-75 kD region contains the following N-terminal amino acid sequence:

AGXLQKNSLLEEVWYLAL

8. A putative protective antigen according to claim 7 further including one or more of the following N-terminal amino acid sequences:

AKNFDFAPSIQGYKKIAHEL

NLKPEQILQLLG

LLKAEXNKXIEEINTXLDN

9. A putative protective antigen according to claim 3 wherein the antigen in the 50-54 kD region contains the following N-terminal amino acid sequence:

MKŁAKLLKGFX(N/L)(M/V)IK
ADP(F/I)(R/E)Y(V/A)PQG(Q/A)X(M/N)VG

10. A putative protective antigen according to claim 3 wherein the antigen in the 52-54 kD region contains the following N-terminal amino acid sequence:

AGXWAKETTKEEKS

11. A putative protective antigen according to claim 10 further including one or more of the following N-terminal amino sequences:

AWVTADGTVN

AIVTADGTVNDNKPNQWVRKY

A putative protective antigen according to claim 3 wherein the antigen in the 45-48 kD region contains the following N-terminal amino acid sequence:

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AGXGQTESGSTSDSKPQAETLKHKV

13. A putative protective antigen according to claim 12 further including one or more of the following internal amino acid sequences:

5 TIYKPDKVLGKVAVEVLRVLIAKKNKASR AEQAITKLKLEGFDTQ KNSQNKIIDLSPEG

14. An isolated nucleic acid fragment encoding a putative protective antigen against <u>Mycoplasma hyopneumoniae</u> or related infections, said nucleic acid fragment including the following sequence, mutants, derivatives, recombinants and fragments thereof:

	10	20	30	40	50	
15	1234567390	1234567890	1234567890	1234567890	1234567890	
	ATGAAAAAA	TGCCACTATA	CCAGAGGAAA	GAGCAGTATA	TAAAATAATT	50
	AAAATTACAT	TITCTTCATT	TGCGCCAGAA	TTTTTAAGAA	TTAGTACATT	100
	AAAAAGTAGA	ACAAAAGTTA	TTAATGTAAA	CATTAGCGCA	ATCCTTAAGA	150
20	AAAAATTAAA	AGTITTATCT	ATTTTTTTA	ATCGAAATCC	AACCAGGCAT	200
	AAATCTTTGT	CAGTATTTAT	CAAGTCGGTA	TITTTICATT	ATTTCTACTA	250
	TATTATTAAA	TGAATTTGCA	TITTCCATAA	TCTAAAATTT	TACATTTTT	300
	TATAACAATT	TITAAAAATT	ACTOTITAAT	TTATAGTATT	TTTTATTT	350
	TTAGTCTAAA	TTATAAATT	ATCTTGAATT	TTATTTGAAT	TTATAATT	- 00
25	TAGTACTAAA	AAATACAAAT	ATTITICCT	ATTCTAAGAA	AAATTCATTT	450
	APPARACITI	ATTGATTTT	ATAGTATAAT	TIGTITGTAT	AATTGAATTA	500
	ACTIGATITG	AAAGGGAACA	AAATGAAAAA	AATGCTTAGA	AAAAAATTCT	5 50
	TGTATTCATC	AGCTATTTAT	GCAACTTCGC	TTGCATCAAT	TATTGCATTT	500
	GTTGCAGCAG	GTTGTGGACA	GACAGAATCA	GGTTCAACTT	CTGATTCTAA	550
30	ACCACAAGCC	GAGACGCTAA	AACATAAAGT	AAGTAATGAT	TCTATTCGAA	700
	TAGCACTAAC	CGATCCGGAT	AATCCTCGAT	GAATTAGTGC	CCAAAAAGAT	750
	ATTATTTCTT	ATGTTGATGA	AACAGAGGCA	GCAACTTCAA	CAATTACAAA	500
	AAACCAGGAT	GCACAAAATA	ACTGACTCAC	TCAGCAAGCT	AATTTAAGCC	850
	CAGCGCCAAA	AGGATTTATT	ATTGCCCCTG	AAAATGGAAG	TGGAGTTGGA	900
35	ACTGCTGTTA	ATACAATTGC	TGATAAAGGA	ATTCCGATTG	TTGCCTATGA	950
	TCGACTAATT	ACTGGATCTG	ATAAATATGA	TTGGTATGTT	TCTTTTGATA	1000
	atgaaaaagt	TGGTGAATTA	CAAGGTCTTT	CACTTGCTGC	GGGTCTATTA	า วิจี0
	GGAAAAGAAG	ATGGTGCTTT	TGATTÇAATT	GATCAAATGA	ATGAATATCT	1100
	AAAATÇACAT	ATGCCCCAAG	AGACAATTTC	TTTTTATACA	ATCGCGGGTT	1150
40	CCCAAGATGA	TAATAATTCC	CAATATTTT	ATAATGGTGC	AATGAAAGTA	7200
	CTTAAAGAAT	Taatgaaaa	TTCGCAAAAT	AAAATAATTG	ATTTATCTCC	1250
	TGAAGGCGAA	AATGCTGTTT	ATGTCCCAGG	ATGAAATTAT	GGAACTGCCG	
	GTCAAAGAAT	CCAATCTTTT	CTAACAATTA	ACAAAGATCC	AGCAGGTGGT	
	AATAAAATCA	AAGCTGTTGG	TICAAAAECA	GCTTCTATTT	TCAAAGGATT	1≈00
45	TETTGECCCA	aatgatggaa	TGGCCGAACA	AGCAATCACC	AAATTAAAAC	1450
	TTGAAGGGTT	TGATACCCAA	+AAATCTTTG	TAACTCGTCA	AGATTATAAT	1500
	GATAAAGSSCA .	AGACTITIAT .	CAAAGĄEGGE	GATCAAAATA	TGACAATTTA	1550

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TAAACCTGAT	AAAGTTTTAG	GAAAAGTTGC	TGTTGAAGTT	CTTCGGGTTT	1500
TAATTGCAAA	GAAAAATAAA	GCATCTAGAT	CAGAAGTCGA	AAACGAACTA	1550
AAAGCAAAAC	TACCAAATAT	TTCATTTAAA	TATGATAATC	AAACATATAA	1700
AGTACAAGGT	AAAAATATTA	ATACAATTIT	AGTAAGTCCA	GTAATTGTTA	1750
CAAAAGCTAA	TGTTGATAAT	CCTGATGCCT	~		1782

15. An isolated nucleic acid fragment according to claim 14 encoding a putative protective antigen wherein the antigen is in the 46-48 kD region including the following nucleic acid sequence, mutants, derivatives, recombinants and fragments thereof:

	10	20	30	40	50	
	1234567890	1234567890	1234557890	1234557890	1234567890	
15	ATGAAAAAA	TGCCACTATA	CCAGAGGAAA	GAGCAGTATA	TAAAATAATT	50
	AAAATTACAT	TITCTTCATT	TGCGCCAGAA	TTTTTAAGAA	TTAGTACATT	100
	AAAAAGTAGA	ACAAAAGTTA	TTAATGTAAA	CATTAGCGCA	ATCCTTAAGA	150
	AAAAATTAAA	AGTTTTATCT	ATTTTTTA	ATCGAAATCC	AACCAGGCAT	200
	AAATCTTTGT	CAGTATITAT	CAAGTEGGTA	TITTICATI	ATTTCTACTA	250
20	AAATATTATT	TGAATTTGCA	TTTTCCATAA	TCTAAAATTT	TACATTITT	300
	TATAACAATT	TITAAAAATT	ACTOTTTAAT	TTATAGTATT	TITATITI	350
	TTAGTCTAAA	TTATAAAATT	ATCTTGAATT	TTATTTGAAT	TTTATAATT	4 00
	TAGTACTAAA	AAATACAAAT	ATTITITCCT	ATTCTAAGAA	AAATTCATTT	450
	TTTAAAAAAA	ATTGATTTTT	ATAGTATAAT	TTGTTTGTAT	AATTGAATTA	5D0
25	ACTIGATITIE	AAAGGGAACA	AAATGAAAAA	AATGCTTAGA	AAAAAATTCT	550
	TGTATTCATC	AGCTATTTAT	GCAACTTCGC	TTGCATCAAT	TATTGCATTT	50 0
	GTTGCAGCAG	GTTGTGGACA	GACAGAATCA	GGTTCAACTT	CTGATTCTAA	550
	ACCACAAGCC	GAGACGCTAA	AACATAAAGT	AAGTAATGAT	TCTATTCGAA	700
	TAGCACTAAC	CGATCCGGAT	AATCCTCGAT	GAATTAGTGC	CCAAAAAGAT	750
30	ATTATTTCTT	ATGTTGATGA	AACAGAGGCA	GCAACTTCAA	CAATTACAAA	30 0
	AAACCAGGAT	GCACAAAATA	ACTGACTCAC	TCAGCAAGCT	AATTTAAGCC	550
	CAGCGCCAAA	AGGATTTATT	ATTGCCCCTG	AAAATGGAAG	TGGAGTTGGA	500
	ACTGCTGTTA	ATACAATTGC	TGATAAAGGA	ATTCCGATTG	TTGCCTATGA	950
	TCGACTAATT	ACTEGATOTE	ATAAATATGA	TTGGTATGTT	TCTTTTGATA	1000
35	ATGAAAAAGT	TGGTGAATTA	CAAGGTCTTT	CACTTGCTGC	GGGTCTATTA	1850
	GGAAAAGAAG	ATGGTGCTTT	TGATTCAATT	GATCAAATGA	ATGAATATCT	1100
	AAAATCACAT	ATGCCCCAAG	AGACAATTTC	TITITATACA	ATCGCGGGTT	1150
	CCCAAGATGA	TAATAATTCC	CAATATTTT	ATAATGGTGC	AATGAAAGTA	1200
	CTTAAAGAAT	TAATGAAAAA	TTCGCAAAAT	AAAATAATTG	ATTTATCTCC	1250
40	TGAAGGCGAA	_	ATGTCCCAGG	ATGAAATTAT	GGAACTGCCG	1300
	GTCAAAGAAT	CCAATCTTTT	CTAACAATTA	ACAAAGATCC	AGCAGGTGGT	1350
	AATAAAATCA	AAGCTGTTGG	TTCAAAACCA	GCTTCTATTT	TCAAAGGATT	1400
	TCTTGCCCCA	AATGATGGAA	TGGCCGAACA	AGCAATCACC	AAATTAAAAC	1450
	TTGAAGGGTT	TGATACCCAA	AAAATCTTTG	TAACTOGTCA	AGATTATAAT	1500
45	GATAAAGCCA	AAACTTTTAT	CAAAGACGGC	GATCAAAATA	TGACAATTTA	1550
	TAAACCTGAT	AAAGTTTTAG	GAAAAGTTGC	TGTTGAAGTT	CTTCGGGTTT	1500
	TAATTGCAAA	GAAAAATAAA	GCATCTAGAT	CAGAAGTOGA	AAACGAACTA	1550
	AAABCAAAAC	TACCARATAT	TTCATTTAAA	TATGATAATC	AAACATATAA	1700
	AGTACAAGGT	ATTATAAAA	ATACAATTIT	AGTAAGTCCA	STAATTGTTA	1750
δC	CAAAAGCTAA	TGTTGATAAT	CCTGATGCCT	AA		1762

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- A method for producing an antibody against a Mycoplasma including 16. providing a biological sample taken a short time after an immune animal has been challenged with a Mycoplasma or Mycoplasma extract taken from the infection site or an area of a lesion or an area close to the infection site or lesion;
- isolating cells from the biological sample; 5 culturing cells in vitro in a suitable culture medium; and harvesting antibodies produced from said cells.
- A method according to claim 16 wherein the biological sample is taken at a 17. predetermined time after the animal has been challenged with a Mycoplasma. 10 preferably 2 to 7 days after challenge.
- A method according to claim 16 wherein the culturing of cells in vitro 18. further includes addition of helper factors to the culture, said helper factors selected from the group including cytokines used alone or in combination. 15 including Interleukin 1, 2, 3, 4, 5, 6, 7 and 8, colony stimulating factors, interferons and any other factors that may be shown to have an enhancing effect on specific B cell secretion.
- A method according to any one of claims 16-18 further including a cell 20 19. activation step including activating the cells isolated to proliferate and secrete and/or release antipodies

said cell activation step including adding a cell activating agent to the culture medium, said ceil activating agent selected from the group including mitogens as herein described and helper factors produced by leukocytes, or their synthetic equivalents or combinations thereof.

- A method according to any one of claims 16-19 wherein the antibody is in 20. the form of the supernatant harvested from the culture medium.
- An antibody against a Mycoplesma prepared according to the method of any one of claims 15-20

GE

22.	A method	of iden	titying a	e putative	protective	antigen	associated	with	3
Mycos	olasma, pre	ferably <u>N</u>	Avcopla	icovń sma	neumoniae,	, said me	thod includi	ng	
	providing								
	2 62	male of	a Muco	nlaema: ai	-d				

a sample of a <u>Mycoplasma;</u> and

an antibody probe including at least one antibody against a Mycoplasma;

probing the Mycoplasma sample with the antibody probe to detect at least one antigen; and

10 isolating the antigen detected.

> 23. A method of purifying a putative protective antigen associated with a Mycoplasma, preferably Mycoplasma hyponeumoniae, said method including providing

15 a crude antigen mixture; and

> an antibody against a Mycoplasma immobilized on a suitable support:

> subjecting the crude antigen mixture to affinity chromatography utilizing the immobilized antipody; and

20 isolating the purified antigen so formed.

- 24 A method for preparing a synthetic antigenic polypeptide against Mycoplasma, preferably Mycoplasma hyppneumoniae, which method includes providing
- 25 a cDNA library or genomic library derived from a sample of bns : smasslccayM

an antibody probe including an antibody prepared according to claim 15;

cenerating synthetic polypeptides from the cDNA library or genomic library. probing the synthetic polypeptides with the antibody probe, and isolating the synthetic antigenic polypeptide detected thereby.

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- 25. A method according to claim 24 wherein the antibody probe includes an antibody raised against an antigen against <u>Mycoplasma hyponeumoniae</u>, or related infections, selected from the group of antigens having approximate molecular weights of 110-114, 90-94, 72-75, 60-64, 52-54 and 46-48 kilodaltons (kD), as herein described, mutants, derivatives and fragments thereof.
- 26. A synthetic putative protective antigen in the 72-75 kD region produced by a method according to claim 24 or 25 having an N-terminal amino acid sequence:

 AGXLQKNSLLEEVWYLAL

27. A synthetic putative protective antigen according to claim 26 further including internal amino acid sequences:

AKNFDFAPSIQGYKKIAHEL NLKPEQILQLLG

15 LLKAEXNKXIEĖINTXLDN

28. A synthetic putative protective antigen in the 60-64 kD region produced by a method according to claim 24 or 25 having an N-terminal amino acid sequence:

MKLAKLLKGFX(N/L)(M/V)IK

ADP(F/I)(R/E)Y(V/A)PQG(Q/A)X(M/N)VG

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29. A synthetic putative protective antigen in the 52-54 kD region produced by a method according to claim 24 or 25 having an n-terminal amino acid sequence:

AGXWAKETTKEEKS

30. A synthetic putative protective antigen according to claim 29 further including internal amino acid sequences.

AWVTADGTVN AIVTADGTVNDNKPNQWVRKY

31. A synthetic putative protective antigen in the 46-48 kD region produced by a method according to claim 24 or 25 having an N-terminal amino acid sequence:

- 33 -

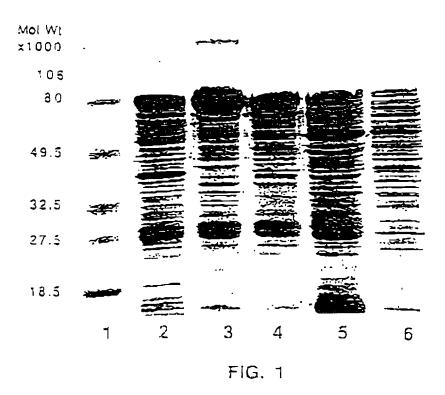
AGXGQTESGSTSDSKPQAETLKHKV

32. A synthetic putative protective antigen according to claim 31 further including internal amino acid sequences:

5 TIYKPDKVLGKVAVEVLRVLIAKKNKASR
AEQAITKLKLEGFDTQ
KNSQNKIIDLSPEG

- 33. A vaccine or veterinary composition including a prophylactically effective amount of at least one putative protective antigen against a <u>Mycoplasma</u> according to any one of claims 1-13.
- 34. A vaccine or veterinary composition according to claim 33 including a plurality of putative protective antigens selected from antigens having approximate molecular weights of 110-114, 90-94, 72-75, 50-64, 52-54 and 46-48 kilodaltons.
 - 35. A vaccine or veterinary composition including an antibody against a Mycoplasma according to claim 21.
 - 36. A diagnostic kit including a diagnostic antigen or fragment thereof according to any one of claims 1-13 and 26-32.
- 37. A method for preventing or treating a <u>Mycoplasma</u> infection, which method including administering to an animal a prophylactically or therapeutically effective amount of at least one putative protective antigen according to any one of claims 1-13.
- 38. An isolated DNA fragment encoding a putative protective antigen against Mycoplasma or related infections, said DNA fragment having a nucleic acid sequence according to Figure 6 or an homologous sequence, and functionally active fragments; mutant, variant or recombinant thereof.

- 39. A clone including a DNA fragment according to claim 38.
- 40. A clone according to claim 39 which is clone pC1-2 as hereinbefore described.
 - 41. An amino acid sequence or functional equivalent thereof encoded by the DNA fragment according to claim 38.
- 10 42. An amino acid sequence or functional equivalent thereof having the amino acid sequence of Figure 7
 - 43. A putative protective antigen or antibody substantially as hereinbefore described with reference to the examples.



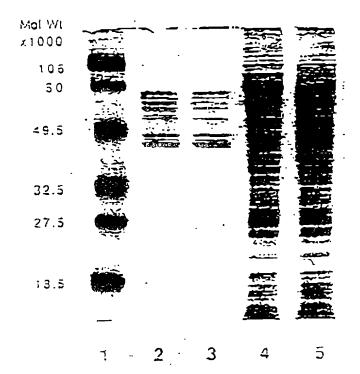
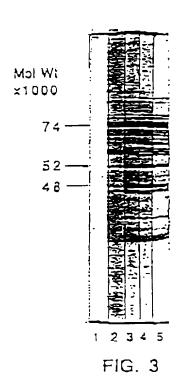
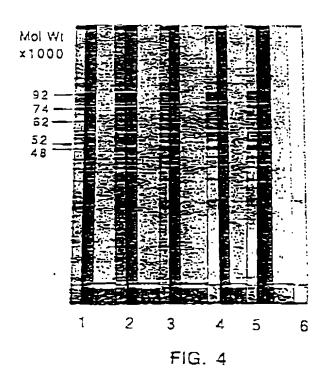


FIG. 2

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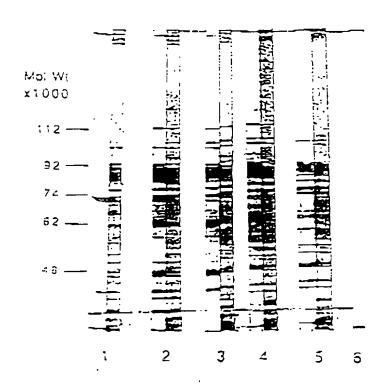


FIG. 5

08/913430

				50	
10	20	30	40	1234557390	
1234567890	1234557890	1234567890	1234547390	1734301390	
					,
71C777777	TGCCACTATA	CCAGAGGAAA	GAGCAGTATA	TALALTARTT	50
AAAATTACAT	TITCTTCATI	TGCGCCAGAA	TTTTTAAGAA	TTAGTACATT	100
ARARAGTAGA	ACARAGITA	TTARTGTARA	G	RICCTIARGR	150
2222277222	AGTTTTATCT	ATTTTTTTTA	ATCGAARTCC	ARCCAGGCAT	200
AAATCTTTGT	CAGTATITAT	CAAGTCGGTA	TITTITCATI	ATTTCTACTA	250
בבבדבדדב ד ד	TG2ATTTGCA	TTTTCCATAA	TCTAAAATTT	TACATTITTT	300
TATARCARTT	TTTARABATT	ACTOTTTAAT	TTATAGTATT	TTTTTATTTT	3 5 0
TTAGTCTARA	TTATALART	ATCTTGARTT	TIRITIGRAN	TTTTATARTT	400
TAGTACTAAA	LINTACARAT	ATTTTTTCCT	ATTOTARGAR	ARATTCATTT	450
777222222	ATTGATTTTT	ATAGTATAAT	TIGITIGIAT	ARTIGRATIA	500
ACTIGATITG	ARREGERACA	<u> 222TGARRR</u>	ARTECTTREA	<u>AAAAA</u> TTCT	550
TGTATTCATC	AGCTATTTAT	GCAACTTCGC	TIGCAICAAT	TATIGCATIT	500
GTTGCAGCAG	GTTGTGGACA	CACAGAATCA	GGTTCARCTT	CIGATICIAA	550 550
ACCACAAGCC	GAGACGCTAA	AACATAAAGT	AAGTAATGAT	TCTATICGAA	700
TAGCACTAAC	CGRICCGGRI	PATCOTOGAT	CARTTAGTGC	CCARARAGAT	750
ATTATTTCTT	ATGITGATGA	<u>AACAGA</u> GGCA	GCAACTTCAA	CAATTACAAA	800
AAACCAGGAT	GCACARARTA	ACTGACTCAC	TCAGCAAGCT	ARTITARGCC	350
CAGCGUCAAA	AGGATTTATT	ATTGCCCCTG	AAAATGGAAG	TGGAGTTGGA	900
ACTGCTGTTA	ATACARTIGO	TGRIRARGEA	ATTCCGATTG	TTGCCTATGA	950
TOGACTARIT	ACTGGATCTG	RTARRIATER	TIGGTATGIT	TCTTTTGATA	1000
RIGRARAGI	TGGTGAATTA	CARGGTCTTT	CACTTGCTGC	GGGTCTATTA	1050
GGARRAGRAG	ATGGTGCTTT	TGATTCAATT	GATCARATGA	ATGRATATOT	1100
HARATERCAT	ATGCCCCAAG	AGRICARTITO	TTTTTATACA	ATCCCCCCTT	1150
CCCRAGATGA	TRATARTICS	CARTATTITT	PIRATESISS	ARTGARAGTA	1200
CTTARAGAAT	TAATGAAAAA	TICGCAAAAT	PRARTARTIG	ATTTATCTCC	1250
TGRAGGGGAA	ARTGOTGTTT	ATGTCCCAGG	RIGARATIAT	GGAACTGCCG	1300
CTCRRAGART	CCARTCITIT	CTARCARTTA	<u>ACAAAGA</u> TCC	AGCAGGTGGT	1350
AATAAAATCA	ARGCTGTTGG	TICAAAACCA	GCTTCTATTT	TCARAGERTT	1400
TOTTGCCCCA	RRIGATOGRA	TGGCCGAACA	AGCARTCACC	RARTTARARC	1450
TTGAAGGGTT	TGATACCCAA	ARRICTITG	TAACTOGTCA	AGRITATART	1500
GATARAGECA	BLACTITIAT	CAAAGACGGC	GATCAAAATA		1550
TARACCTGAT	AAAGTTTTAG	GRARAGTTGC	TGTTGRAGIT	CTICGGGIII	1500
TARTTGCARA	CALLATARA		CAGAAGTCGA		1550
AAAGCAAAAC	TACCAAATAT	TTCATTTAAA	TATGATARIC	AAACATATAA	1700
AGTACAAGGT		ATACAATTT	<u>AGTARGTOCA</u>	STRAFFSTTA	1750
	TOTTONIN	CCTGATGCCT	22		<u> </u>

1	20	30	40	50	
1234567890	1234557890	1234557890	1234557890	1234567890	
				25/202255	.
	YSSAIYATSL				50
HKV5NDSIRI	ALTDPDNPRW	ISAQXDIISY	VDETERATS?	ITENQDAQNN	100
WLTQQANL5?	APKGFIIAPE	NGSGVGTAVN	TIADKGIPIV	AYDRLITGSD	150
KYDWYVSFDN	EKVGELQGLS	LAAGLLGKED	CFID2IDOW	EATK2#34565	200
TISFYTIAGS	QDDNNSQYFY	NEWKYLKEL	VECASONKIID	LSPECENAVY	250
VPGWNYGTAG	QRIQSFLTIN	XDPAGGNXIX	AVGSKPASIT	KGFLAPNDGM	300
AEQAITKLKL	EGFDIQKIFV	TRODYNDKAK	TFIKDGDQNM	TIYK?DKVLG	350
KVAVEVLRVL	IAKKOKASRS	EVENELKAKL	PHISEKYDNQ	TYKVQGKNIN	400
TILVSPVIVI	KANVDNPDA				419

Attorney's Docket No. U 011415-0	PATENT
COMBINED DECLARATION AND POWER (OF ATTORNEY
(ORIGINAL, DESIGN, NATIONAL STAGE OF PCT, SUPPL CONTINUATION OR C-I-P)	EMENTAL, DIVISIONAL,
As a below named inventor, I hereby declare that:	
TYPE OF DECLARATION	
This declaration is of the following type: (check one applicab	ole item below)
□ original	
☐ design	
☐ supplemental	
NOTE: If the declaration is for an International Application being file continuation-in-part application, do not check next item; check as	d as a divisional, continuation of oppopnate one of last three items.
NOTE: If one of the following 3 items apply, then complete and also attach CONTINUATION OR C-I-P.	h ADDED PAGES FOR DIVISIONAL,
☐ divisional	
continuation	
□ continuation-in-part (C-I-P)	
INVENTORSHIP IDENTIFICATION	ON
WARNING: If the inventors are each not the inventors of all the claims, a the ownership of all the claims at the time the last claims submitted.	n explanation of the facts, including ed invention was made, should be
My residence, post office address and citizenship are as start believe I am the original, first and sole inventor (if only one original, first and joint inventor (if plural names are listed below is claimed and for which a patent is sought on the invention	name is listed below) or ar) of the subject matter which
TITLE OF INVENTION	
ANTIGEN COMPOSITION AGAINST MYCOPLASMA	
SPECIFICATION IDENTIFICATE	ON
the specification of which: (complete (a), (b) or (c))	
(a) is attached hereto.	
(b) was filed on as as	Serial No. 0 /
or Express Mail No., as Serial No. not yet know and was amended on	wn
(Deslamtion and Payer of A	Namey [1-1]nage of :

NOTE:	Amendments filed after the original papers are deposited with the PTO which contain new matter are not accorded a filing date by being referred to in the declaration. Accordingly, the amendments involved are those filed with the application papers or, in the case of a supplemental declaration, are those amendments claiming matter not encompassed in the original statement of invention or claims. See 37 CFR 1.67.
(c) 🛭	PCT/AU96/00149 filed on 15 March 1996 and as
	amended under PCT Article 19 on (if any).
ACK	NOWLEDGEMENT OF REVIEW OF PAPERS AND DUTY OF CANDOR
	by state that I have reviewed and understand the contents of the above identified ation, including the claims, as amended by any amendment referred to above.
l ackr	nowledge the duty to disclose information
2	which is material to patentability as defined in 37, Code of Federal Regulations, § 1.56
	(also check the following items, if desired)
	and which is material to the examination of this application, namely, information where there is a substantial likelihood that a reasonable examiner would consider it important in deciding whether to allow the application to issue as a patent, and
	☐ In compliance with this duty there is attached an information disclosure statement in accordance with 37 CFR 1.98.
	PRIORITY CLAIM (35 U.S.C. § 119)
foreign applicat below a certifica the Unit	by claim foreign priority benefits under Title 35, United States Code, § 119 of any application(s) for patent or inventor's certificate or of any PCT international ion(s) designating at least one country other than the United States of America listed and have also identified below any foreign application(s) for patent or inventor's te or any PCT international application(s) designating at least one country other than ted States of America filed by me on the same subject matter having a filing date that of the application(s) of which priority is claimed.
	(complete (d) or (e))
(d) [no such applications have been filed.
(e) 🛚	such applications have been filed as follows.
NOTE:	Where item (c) is entered above and the International Application which designated the U.S. itself claimed priority check item (e), enter the details below and make the priority claim.

A. PRIOR FOREIGN/PCT APPLICATION(S) FILED WITHIN 12 MONTHS (6 MONTHS FOR DESIGN) PRIOR TO THIS APPLICATION AND ANY PRIORITY CLAIMS UNDER 35 U.S.C. § 119

COUNTRY (OR INDICATE IF PCT)	APPLICATION NUMBER	DATE OF FILING (day, month, year)	PRIORITY CLAIMED UNDER 37 USC 119
AU	PN 1789	16 March 1995	☑ YES NO □
PCT	PCT/AU96/00149	15 March 1996	☑ YES NO □
			☐ YES NO ☐
			☐ YES NO ☐
			☐ YES NO ☐

ALL	FOREIGN APPLICATION(S), IF ANY FILED MORE THAN 12 MONTHS (6 MONTHS FOR DESIGN) PRIOR TO THIS U.S. APPLICATION	
		_

NOTE: If the application filed more than 12 months from the filing date of this application is a PCT filing forming the basis for this application entering the United States as (1) the national stage, or (2) a continuation. divisional, or continuation-in-part, then also complete ADDED PAGES TO COMBINED DECLARATION AND POWER OF ATTORNEY FOR DIVISIONAL, CONTINUATION OR C-I-P APPLICATION for benefit of the prior U.S. or PCT application(s) under 35 U.S.C. § 120.

POWER OF ATTORNEY

I hereby appoint the following attorney(s) and/or agent(s) to prosecute this application and transact all business in the Patent and Trademark Office connected therewith. (List name and registration number)

PAUL B. WEST, 18947 JOSEPH H. HANDELMAN, 26179 JOHN RICHARDS, 31053 JOHN J. CRYSTAL, 26360-RICHARD J. STREIT, 25765 ALAN K. ROBERTS, 17777-S. DELVALLE GOLDSMITH, 14383

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(check the following item, if applicable)

Attached as part of this declaration and power of attorney is the authorization			
of the above-named attorney(s) to accept and follow instructions from my			
representative(s).			

(Declaration and Power of Attorney [1-1]—page 3 of 5)

SEND CORRESPONDENCE TO

DIRECT TELEPHONE CALLS TO: (Name and telephone number)

LADAS & PARRY 26 WEST 61ST STREET NEW YORK, NEW YORK 10023

Eull name of cale or first inventor

Full name of second joint inventor, if any

(212)708-1930

DECLARATION

I hereby declare that all statements made herein of my own knowledge are true and that all statements made on information and belief are believed to be true; and further that these statements were made with the knowledge that willful false statements and the like so made are punishable by fine or imprisonment, or both, under Section 1001 of Title 18 of the United States Code, and that such willful false statements may jeopardize the validity of the application or any patent issued thereon.

SIGNATURE(S)

NOTE: Carefully indicate the family (or last) name as it should appear on the filing receipt and all other documents.

John-	The control of the co	WALKER	
(GIVEN NAME)	(MIDDLE INITIAL OR NAME)	. FAMILY (OR LAST NAME)	
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Date 6/10/97	Country of Citizenship		
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Post Office Address 26	Clapham Street, Balwyn, <u>Vic</u> t	toria 3103 Australia	
Fost Office Address		Lul	

Rogan FAMILY (OR LAST NAME) (MIDDLE INITIAL OR NAME) (GIVEN NAME) Inventor's signature Date 6 October 1997 Country of Citizenship 73 Greenford Street, Chapel Hill, Queensland 4069 Australia

Post Office Address 73 Greenford Street, Chapel Hill,

(Declaration and Power of Attorney [1-1]—page 4 of 5)

Full name	e of third join	t inventor, if any	
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	s signature _	70.907.7	Australia
	25/9/97	Country of Citizenship	
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Post Offi	ce Address _	1A Diana Drive, Blackburn, N	/ictoria 3130 Australia
	 		<u> </u>
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(Declaration and Power of Attorney [1-1]—page 5 of 5)